



The University of Manchester

The University of Manchester Research

# Deforestation caused abrupt shift in Great Lakes nitrogen cycle

DOI:

10.1002/lno.11428

**Document Version** 

Accepted author manuscript

Link to publication record in Manchester Research Explorer

Citation for published version (APA):
Guiry, E. J., Buckley, M., Orchard, T. J., Hawkins, A. L., Needs-howarth, S., Holm, E., & Szpak, P. (2020).
Deforestation caused abrupt shift in Great Lakes nitrogen cycle. *Limnology and Oceanography*. https://doi.org/10.1002/lno.11428

Published in:

Limnology and Oceanography

Citing this paper

Please note that where the full-text provided on Manchester Research Explorer is the Author Accepted Manuscript or Proof version this may differ from the final Published version. If citing, it is advised that you check and use the publisher's definitive version.

**General rights** 

Copyright and moral rights for the publications made accessible in the Research Explorer are retained by the authors and/or other copyright owners and it is a condition of accessing publications that users recognise and abide by the legal requirements associated with these rights.

Takedown policy

If you believe that this document breaches copyright please refer to the University of Manchester's Takedown Procedures [http://man.ac.uk/04Y6Bo] or contact uml.scholarlycommunications@manchester.ac.uk providing relevant details, so we can investigate your claim.



### Deforestation caused abrupt shift in Great Lakes nitrogen cycle

- <sup>\*1,2</sup>Eric J. Guiry, <sup>3</sup>Michael Buckley, <sup>4</sup>Trevor J. Orchard, <sup>5</sup>Alicia L. Hawkins, <sup>6,7,8</sup>Suzanne Needs-
- 3 Howarth, <sup>9</sup>Erling Holm, <sup>1</sup>Paul Szpak

4

1

- <sup>1</sup> Department of Anthropology, Trent University, 1600 West Bank Dr., Peterborough, ON, K9L 0G2, Canada
- 6 (eguiry@lakeheadu.ca; paulszpak@trentu.ca)
- <sup>2</sup> School of Archaeology and Ancient History, University of Leicester, Mayor's Walk, Leicester, LE1 7RH, United
- 8 Kingdom
- 9 <sup>3</sup> Department of Earth and Environmental Sciences, Manchester Institute of Biotechnology, The University of
- Manchester, 131 Princess Street, Manchester M1 7DN, United Kingdom (m.buckley@manchester.ac.uk)
- <sup>4</sup> Department of Anthropology, University of Toronto Mississauga, 3359 Mississauga Road, Mississauga, ON L5L
- 12 1C6, Canada (trevor.orchard@utoronto.ca)
- <sup>5</sup> Archaeology program, School of the Environment, Laurentian University, 935 Ramsey Lake Rd., Sudbury, ON
- 14 P3E 2C6, Canada (ahawkins@laurentian.ca)
- <sup>6</sup> Perca Zooarchaeological Research, Toronto, Canada (suzanne.needs.howarth@utoronto.ca)
- <sup>7</sup> The Archaeology Centre, University of Toronto, 19 Russell Street, Toronto, ON M5S 2S2, Canada
- <sup>8</sup> Trent University Archaeological Research Centre, 1600 West Bank Dr., Peterborough, ON K9L 0G2, Canada
- <sup>9</sup> Department of Natural History, Royal Ontario Museum, 100 Queen's Park, Toronto, ON M5S 2C6, Canada
- 19 (erlingh@rom.on.ca)

20

21

#### Abstract

- 22 Despite the longstanding significance of North America's Great Lakes, little is known about their
- pre-industrial ecology. Here, we report on when and how humans first became a main driver of
- Lake Ontario's nutrient dynamics. Nitrogen isotope analyses of archaeological fish show that,
- 25 prior to the 1830s, Lake Ontario's nitrogen cycle and the trophic ecology of its top predators had
- remained stable for at least 800 years, despite Indigenous and historical European agricultural
- 27 land management across the region. An abrupt shift in the nitrogen isotope composition of Lake
- Ontario's fish community is evident in the early to mid-nineteenth century and reflects the
- 29 initiation of industrial-scale forest clearance. These data show how the nitrogenous nutrient
- regimes of even the world's largest freshwater ecosystems can be highly sensitive to short-term
- 31 watershed forest cover disturbances and indicate a profound shift in the relationship between
- 32 humans and their environment.

33

34

#### **Keywords**

35 Historical Ecology, Archaeology, Great Lakes, Nitrogen Cycle, Fish, Deforestation

#### Introduction

37

38

39

40

41

42

43

44

45

46

47

48

49

50

51

52

53

54

55

56

57

58

59

freshwater environments have profoundly altered aquatic biogeochemical cycles and broader nutrient dynamics, resulting in widespread ecosystem dysfunction, loss of biodiversity, and degraded water quality (Smith and Schindler 2009). While numerous studies have documented how increasing anthropogenic nitrogen inputs are incorporated into aquatic food webs, promoting cultural eutrophication and altering trophic dynamics in the recent past (Schindler, et al. 2006), less consideration has been given to how historical and preindustrial populations have impacted aquatic ecosystems. In the context of recent debate about the timing of the origins of the Anthropocene (Lewis and Maslin 2015), in which past societies are considered as potential architects of the first broad-scale environmental changes, the role of humans as drivers of biogeochemical processes such as the nitrogen cycle is becoming increasingly important (Guiry, et al. 2018; Hadley, et al. 2010; Kintigh, et al. 2014). For conservation and restoration efforts in freshwater ecosystems, where cultural eutrophication caused by increased nutrient loading poses one of the most significant threats globally (Smith and Schindler 2009), long-term retrospectives that document when and how human activities first began to alter natural ecosystem processes, and the nitrogen cycle in particular, could provide a valuable framework for evaluating which modern human activities pose the greatest risks (Canfield, et al. 2010; Humphries and Winemiller 2009). Paleolimnological proxy indicators based on the physical, biological, chemical, and isotopic compositions of sediments have long been used to establish trends in past aquatic productivity and water quality (Beeton 1965; Hodell and Schelske 1998; Jeffers, et al. 2015; Schelske, et al. 1983), but, because of potential taphonomic issues (Anderson 2014; Lu, et al. 2014) and because these approaches

Throughout the twentieth century, increased anthropogenic contributions of reactive nitrogen to

usually do not incorporate consumers, they are unable to measure the direct impacts of anthropogenic nitrogen loading on nutrient dynamics in the wider biotic community (e.g., invertebrates, fish, birds; although for a growing literature on invertebrate analyses see Anas, et al. 2019; Frossard, et al. 2014; Perga 2010; Perga, et al. 2010; Schilder, et al. 2017; van Hardenbroek, et al. 2010). Moreover, while many studies using isotopic analyses of museumarchived vertebrate tissues have been able to make important contributions to understanding how biotic communities have responded to human-caused environmental change during the twentieth century (Fera, et al. 2017), a lack of suitable specimens from early historical and pre-industrial time periods has, in most cases, prevented analyses of longer-term environmental variation in vertebrate taxa (Szpak, et al. 2018). In this context, stable nitrogen isotope and other analyses of ancient fish remains from archaeological repositories can provide an invaluable opportunity to gain direct insight into how food web and nutrient dynamics within pre-industrial ecosystems functioned and responded to impacts from human activities at varying temporal and spatial scales (Guiry, et al. 2016a). Using isotopic compositions of fish bone or scales has some interpretive advantages relative to more commonly analyzed materials, such as bulk sedimentary organic matter, because fish stable nitrogen isotope composition is linked firmly with a known ecological point (i.e. a particular species with well-understood ecology and trophic affinity) and spatiotemporal framework (Trueman and Moore 2007) and can be easily parsed to remove diagenetically altered samples using robust quality-control criteria (DeNiro 1985; Szpak 2011). This is important because, in contrast to analyses of bulk sedimentary organic matter, which is composed of a spatially, temporally, and biologically heterogeneous mixture of sources of detritus (Lu, et al. 2014), with fish bone or scales, the ability to compare "taxonomically anchored" isotopic patterns across

60

61

62

63

64

65

66

67

68

69

70

71

72

73

74

75

76

77

78

79

80

81

different parts of a food web will enable much higher-resolution interpretations of environmental change at both the level of nutrient dynamics and the level of broader food web structures. Moreover, with respect to measuring the isotopic compositions of ancient organic materials, the ability to purify collagen extracted from bone (through pretreatment steps outlined below) and assess the extent to which its constituent carbon and nitrogen are biogenic (e.g., using wellestablished and precise C:N<sub>Atomic</sub> criteria (DeNiro 1985; Szpak 2011) provides an additional advantage relative to other important paleontological materials (e.g., chitinous remains from sub fossil invertebrates; Anas, et al. 2019) and makes bone and scale collagen particularly well suited for inter-site comparisons of isotopic variation in biota across time and space. Moreover, because collagen from bones and scales is constructed and remodeled slowly throughout the life of an organism, the isotopic composition of collagen can provide a lifetime average record of dietary intake and environmental conditions that is less susceptible to seasonal or short-term idiosyncratic shifts in behavior (Hobson and Clark 1992). Bone collagen is therefore well suited for reconstructing long-term trends in biology and ecology (Bump, et al. 2007) and can provide an additional perspective with potential to complement and build on the success of previous, well-established paleolimnological proxy indicators (e.g., physical, chemical, and isotopic compositions of sediments and invertebrates; Beeton 1965; Hodell and Schelske 1998; Jeffers, et al. 2015; Perga 2010; Schelske, et al. 1983). We use stable nitrogen isotope compositions and peptide mass fingerprinting (Zooarchaeology by Mass Spectrometry, hereafter, ZooMS) of late Holocene (900-2015 CE) fish bone collagen from a large number of sites associated with Lake Ontario (Figure 1), the most easterly of the Laurentian Great Lakes (hereafter, Great Lakes), to document long-term trends in regional biogeochemical cycles. Results show that, in comparison with the industrial era, the nitrogen

83

84

85

86

87

88

89

90

91

92

93

94

95

96

97

98

99

100

101

102

103

104

cycle and trophic structure of this Great Lakes ecosystem remained remarkably stable until the 1830s, despite millennia of Indigenous agricultural and other land management, decades of European settlement, and climatic fluctuations. After this time, increased N-loading from forestry- and agriculture-induced soil erosion caused an unprecedented and abrupt bottom-up shift throughout the entire aquatic ecosystem of Lake Ontario.

#### Context

106

107

108

109

110

111

112

113

114

115

116

117

118

119

120

121

122

123

124

125

126

127

128

With more than 20% of the world's surface freshwater, North America's Great Lakes are of tremendous ecological, economic, and social interest (Sterner, et al. 2017). Since monitoring programs began, the Great Lakes have experienced significant ecological and chemical changes in response to pollution, hydrological controls, and species invasion (Allan, et al. 2013; Stewart, et al. 2016). Paleolimnological studies show that recent industrial activities have had profound impacts on Great Lakes productivity (Beeton 1965; Hodell and Schelske 1998; Jeffers, et al. 2015; Meyers 2003; Schelske and Hodell 1991b; Schelske and Hodell 1995; Schelske, et al. 1988; Schelske, et al. 1983), especially through increased phosphorus and nitrogen loading that led to eutrophication of Lake Ontario and Lake Erie by the twentieth century (Hodell and Schelske 1998; Lu, et al. 2010). With respect to nutrient dynamics, particularly the nitrogen cycle, which plays a vital role in primary production in naturally oligotrophic ecosystems such as Lake Ontario (Lean and Knowles 1987; Leggett, et al. 2000), the extent to which these changes represent a departure from longer-term trends in broader food web structure and nutrient dynamics remains unclear (Sterner, et al. 2017; Supplementary Information [SI] 1). But this extent could be determined through isotopic reconstructions of past nutrient and food web dynamics using historical or archaeological fish specimens, as these can provide a complementary isotopic record for ancient environmental variation.

Human activities have modified the cycling of nitrogen in aquatic environments across the globe, contributing to the ongoing threat of cultural eutrophication (Gruber and Galloway 2008; Smith and Schindler 2009). The majority of these impacts occur when anthropogenic nitrogen inputs and/or chemical or physical changes to an environment alter the conditions regulating the nitrogen cycle, thereby disrupting the flow of nitrogenous nutrients through an ecosystem (Kendall, et al. 2007). Modeling how nitrogen moves into and through aquatic ecosystems is, therefore, of considerable importance for conservation efforts seeking to restore heavily impacted watersheds (Denk, et al. 2017). With respect to broad-scale human impacts, such factors as climate change, acid rain, nutrient loading, and invasive species introductions can significantly alter the biogeochemical processes that balance an aquatic ecosystem's nitrogen cycle and often, in turn, cause detectable changes in the isotopic composition of nitrogen pools that are available to support aquatic life (Anderson and Cabana 2005; Botrel, et al. 2014; Lake, et al. 2001). Stable nitrogen isotopic compositions of consumer tissues show a stepwise increase, by roughly 3-4%, in  $\delta^{15}$ N values with each trophic level step up in a food web and have therefore traditionally provided a powerful indicator for food web interactions in ecology and archaeology (Post 2002). However, because any impact on the  $\delta^{15}N$  of aquatic nitrogen pools at the base of a food web (i.e., in various pools of dissolved inorganic nitrogen [DIN]) is passed up the trophic ladder to producers (phytoplankton) and their consumers (invertebrates and fish), isotopic analyses of tissues from aquatic producers and consumers can also serve as a highly integrative indicator for anthropogenic impacts on nutrient dynamics, in addition to food web structure, in aquatic environments (Hoffman, et al. 2012; Morrissey, et al. 2013). While a majority of isotopic research has approached the question of changing freshwater nutrient dynamics through analyses of sedimentary organic matter (e.g., Dubois, et al. 2018; Talbot

129

130

131

132

133

134

135

136

137

138

139

140

141

142

143

144

145

146

147

148

149

150

2001), a growing number of studies are demonstrating that  $\delta^{15}N$  compositions of invertebrates 152 and fish can provide a highly sensitive record for environmental change (e.g., Anas, et al. 2019; 153 Fera, et al. 2017; Lumb and Johnson 2012; Perga, et al. 2010). 154 The natural abundance of <sup>15</sup>N in DIN is largely controlled by three factors (for reviews see, 155 Finlay and Kendall 2007; Guiry 2019; Kendall 1998; Kendall, et al. 2007; Talbot 2001): 156 productivity (Hodell and Schelske 1998; Schelske and Hodell 1991a), environmental conditions 157 (temperature, oxygenation, and pH; Finlay and Kendall 2007; Knowles 1982), and nitrogen 158 inputs (quantity and isotopic composition; Heaton 1986; Lake, et al. 2001). These factors are 159 necessarily interrelated because, for instance, increasing the input of reactive nitrogen can 160 161 increase productivity, and this can affect environmental conditions that regulate other parts of the nitrogen cycle (Finlay and Kendall 2007). Moreover, nitrogen-limitation rates can influence 162 plankton community composition, which is an integral component of determining the presence 163 of nitrogen-fixing bacteria (Gu 2009) that can further modify the isotopic composition of an 164 aquatic ecosystem by introducing isotopically distinctive atmospheric nitrogen. Human activity 165 should affect those factors controlling the nitrogen cycle and its isotopic composition (for review 166 see Guiry 2019) by: 1) contributing reactive nitrogen (i.e., through inputs directly from sewage, 167 agriculture, and soil erosion – typically, but not always, leading to elevated biota  $\delta^{15}$ N (Anderson 168 and Cabana 2005; Morrissey, et al. 2013)), which itself leads to increasing productivity (creating 169 greater nitrogen demand and thereby possibly promoting the importance of N-fixing algae -170 leading to higher and lower biota  $\delta^{15}$ N, respectively(Gu 2009; Pennock, et al. 1996)), and 2) 171 172 changing chemical conditions, such as pH and oxygen levels, that regulate transformations 173 between important forms of reactive nitrogen (altering the dynamics for nitrification and

denitrification – with potential to push biota  $\delta^{15}$ N higher or lower (Collister and Hayes 1991; Sebilo, et al. 2006)).

174

175

176

177

178

179

180

181

182

183

184

185

186

187

188

189

190

191

192

193

194

195

196

With the longest history of European and Indigenous agricultural land management and settlement among the Great Lakes, Lake Ontario and its watershed are an ideal context in which to investigate how large-scale human activities can impact major freshwater environments over time (Hodell and Schelske 1998). Because Lake Ontario receives a large amount of its water from land runoff, it is particularly sensitive to broader ecological processes and human disturbances occurring across the watershed, particularly in terrestrial and riparian zones. Moreover, the significant and highly visible environmental degradation of Lake Ontario's watershed throughout the twentieth century, as well as its close proximity to major population centers, has spurred considerable research into its ecological structure (e.g., Bogue 2001; Estepp and Reavie 2015; Meyers 2003; Stewart, et al. 2016). For this reason, the lake has a relatively well-documented ecological and biogeochemical history, with numerous isotopic studies of modern fish identifying recent trends in community structure, trophic dynamics, and energy flows (Colborne, et al. 2016; Fera, et al. 2017; Kiriluk, et al. 1995; Lumb and Johnson 2012; Rush, et al. 2012; Yuille, et al. 2015). The longest-term of these studies, however, only offers a 70-year retrospective, one that postdates intensification of European settlement of Lake Ontario's watershed by more than 150 years.

While a number of important isotopic studies have helped to document longer-term variability in Lake Ontario's nitrogenous nutrient dynamics (SI 1), these have focused on isotopic compositions of organic matter in lake sediments rather than specific taxa. With respect to the last millennium, the timeframe over which human land management would have intensified, most (95%) of these analyses focus on the period after the mid-nineteenth century and show

substantial change in the nitrogen isotopic composition of organic matter settling out of Lake Ontario's water column (SI 1; Figure S1, Figure 2; Hodell and Schelske 1998; McFadden, et al. 2004). Only a small fraction (5%; n=8 of 178) of analyses cover the early historical period (1600s-1850 CE), when European activities would have started impacting the region, and precontact period, when Indigenous farming and other land management activities were accelerating across the watershed (i.e., starting with the Late Woodland, c. 900 CE). Moreover, only one lake sediment isotopic dataset spans both time frames and is not consistent with the others (SI 1, Figure 2). While these data provide an invaluable macro-scale perspective on general patterns in nutrient dynamics in Lake Ontario, they cannot be used to assess the consequences of shifting nitrogenous nutrient regimes for the broader food web (invertebrates and fish). Moreover, the coarse temporal resolution of the lake sediment isotopic record prior to 1850 prevents detailed analyses of when the human activities began to impact the nitrogen cycle and to what extent early historical and Indigenous activities may have been important drivers for change. In this context, isotopic analyses of archaeological fish can provide a complementary line of evidence for nutrient dynamics and food web structure.

#### Materials and methods

197

198

199

200

201

202

203

204

205

206

207

208

209

210

211

212

213

214

215

216

217

218

219

Sample description: Sampling focused on three taxa in the family Salmonidae: lake trout (Salvelinus namaycush; n=222), Atlantic salmon (Salmo salar; n=158), and whitefishes (Coregonus spp.; primarily C. clupeaformis and C. artedi, see below; n=277; SI 2, Table S1). These taxa were selected to provide a highly integrated record of changes to Lake Ontario's nitrogen cycle and trophic structure. The upper trophic position and longer lifespan (5–15 years) as well as pelagic foraging and highly mobile behavior of these species means that their diets will incorporate a broad range of resources from across the lake (Fera, et al. 2017; Guiry, et al.

2016a; Rush, et al. 2012), providing a time-averaged perspective on processes affecting the nitrogen cycle at a lake-wide, multi-seasonal scale. A comparison of isotopic variation among three taxa, each with a different behavioral strategy and trophic position (Fera, et al. 2017; Holm, et al. 2009; Mumby, et al. 2018), should make isotopic trends associated with dietary shifts at the species level (as opposed to changes at the base of the food web) clearly discernible and, therefore, these data also give a long-term perspective on the stability of Lake Ontario's food web structure.

220

221

222

223

224

225

226

227

228

229

230

231

232

233

234

235

236

237

238

239

240

241

242

With the exception of nine Atlantic salmon samples, all archaeological specimens are from sites within the Lake Ontario watershed. As Lake Ontario is the only known source of Atlantic salmon, specimens from this species at sites outside the watershed were also included because they represent individuals originating from Lake Ontario. Where possible, archaeological fish bone specimens were selected based on minimum number of individual counts per archaeological context in order to avoid sampling the same individual multiple times. For all species, archaeological sampling efforts targeted bone specimens from adult-sized fish in order to exclude juveniles, who may feed at a lower trophic level. All specimens were examined by one or more of three zooarchaeologists (S.N.-H., A.H., T.O.) with specific expertise in the identification through comparative osteology of archaeological fish bones from relevant taxa in the Great Lakes region (Hawkins, et al. 2019). The archaeological samples are largely derived from fish vertebra, because they are more ubiquitous than fish cranial and pectoral girdle bones in the archaeological collections. Zooarchaeologists determined species for each specimen where possible, based on clear morphological differences between taxa. Due to overlaps in osteology and the possibility of hybrids, morphological examination of *Coregonus* vertebrae and many Coregonus cranial and pectoral girdle bones cannot be confidently used to assign species-level identification in the Lake Ontario watershed. However, of the five Coregonus species that are native to Lake Ontario, only two, lake whitefish (C. clupeaformis) and cisco (C. artedi), inhabit waters shallow enough to have been broadly accessible using fishing technologies (birch bark canoes and gill nets) available in the pre-contact past, and it is therefore highly likely that archaeological Coregonus of that time are composed of these taxa. Some bones in the genera Salvelinus and Salmo are also morphologically similar to each other, resulting in additional higher-level taxonomic identifications (including to Salmo salar/Salvelinus namaycush and Salmonidae). In cases where a species-level taxonomic identification was not achieved with a high degree of certainty, we undertook ZooMS analyses (n=292) to confirm identifications, comparing the archaeological samples with reference sequences we generated for S. salar, S. namaycush, brook trout (Salvelinus fontinalis), round whitefish (Prosopium cylindraceum), C. clupeaformis, and *C. artedi* (Supporting Information 4 Table S1). Dating for pre-contact specimens is based on published (SI 2, Table S2) radiocarbon dates, ceramic seriations, and reconstructed village occupation sequences (e.g., Williamson 2014), to occupations within 100-year bins. Excellent chronological control for data from Indigenous archaeological sites reflects the highly standardized lifecycle in which settlements were built, occupied, decommissioned, and left behind in favor of relocation every 10-30 years (Warrick 2008). The relative dearth of data from the seventeenth and eighteenth centuries reflects changes in Indigenous settlement patterns during this period. Reduced sample sizes in the nineteenth century reflect lower frequencies for fish bone at Euro-Canadian archaeological sites. Historical fish scale samples of adult specimens of known date were taken from museum-archived collections with assistance from an Ontario fish identification expert (E.H.). Analyses of scales from museum-archived Coregonus specimens include both C. clupeaformis and C. artedi

243

244

245

246

247

248

249

250

251

252

253

254

255

256

257

258

259

260

261

262

263

264

because: 1) these two species likely account for the majority of archaeological specimens and will therefore be directly comparable; and 2), based on  $\delta^{15}$ N values, they appear to have fed at the same trophic level in early twentieth-century Lake Ontario. For data sourced from the literature (n=834; (Colborne, et al. 2016; Fera, et al. 2017; Guiry, et al. 2016a; Kiriluk, et al. 1995; Rush, et al. 2012; van der Merwe, et al. 2003; Yuille, et al. 2015)), fork length, when available, was used to exclude juveniles, which may feed at a lower trophic level. Sample preparation: Samples weighing between 30 and 300 mg were cut from bone specimens, residual bone lipids were removed with a series of 2:1 chloroform:methanol ultrasonic baths (solution refreshed every 10 minutes until visible signs of reaction ceased), and the samples were then left to air dry (Guiry, et al. 2016b). Bone samples were then demineralized in 0.5 M hydrochloric acid (HCl) for several days at 4°C and then rinsed to neutrality in Milli-Q water. Humic acids and other base soluble contaminants from the burial environment were removed from demineralized bone samples with successive treatments in 0.1 M sodium hydroxide in an ultrasonic bath (solution refreshed every 15 minutes until visible signs of reactions ceased) and then rinsed to neutrality in Milli-Q water (Szpak, et al. 2017a). Bone samples were then solubilized in  $10^{-3}$  M HCl in an oven at  $65^{\circ}$ C for 48 hours. The resulting solutions were centrifuged, after which the solubilized collagen fraction was transferred to a new tube and then frozen and lyophilized. The protein fraction of fish scale is also composed primarily of collagen, which is compositionally and isotopically comparable to fish bone collagen (Guiry, et al. 2016a; Trueman and Moore 2007). Fish scales samples underwent a succession of three rinses in Milli-Q water in an ultrasonic bath for five minutes each (Guiry, et al. 2016a). Scale samples were then soaked for five minutes in 1.0 M HCl in an ultrasonic bath to remove the mineral phase of the scale's

266

267

268

269

270

271

272

273

274

275

276

277

278

279

280

281

282

283

284

285

286

287

external plate as well as any microscopic debris that may have become adhered to the external surfaces of the fish specimens during preservation and storage. Scale samples were then rinsed to neutrality in Milli-Q water in an ultrasonic bath and left to air dry. The historical fish specimens had been preserved using formalin fixation and then stored in ethanol. While this form of preservation is known to introduce small quantities of carbon, which can slightly alter the stable carbon isotope composition of biological tissues, formalin and ethanol provide no new sources of nitrogen and therefore have a minimal influence the nitrogen isotopic composition of preserved fish scales. To confirm this, we also compared the percent carbon-to-nitrogen ratio (C:N) of historical scales with that of modern fish scales (see below; (Szpak 2011)) to establish that diagenetic nitrogen is not present. Isotopic analyses: The stable nitrogen isotope composition as well as percent carbon and nitrogen were measured on 0.5 mg samples of bone and scale collagen using an Elementar Vario MICRO cube elemental analyzer coupled via continuous flow to an Isoprime isotope ratio mass spectrometer in the Archaeology Chemistry Laboratory at the University of British Columbia, Canada. Duplicate or triplicate analyses were performed on 14% of samples. For a full account of procedures used for calibration of isotopic values as well as isotopic uncertainty calculations see SI 3. Analytical uncertainty for  $\delta^{15}N$  measurements was +0.20% (Szpak, et al. 2017b; SI 3). The following data quality criteria were applied for establishing the integrity of  $\delta^{15}N$ measurements from bone and scale collagen (DeNiro 1985): 1) % carbon and % nitrogen values above 13% and 4.8%, respectively; and 2) atomic C:N values falling between 2.9 and 3.6. ZooMS analyses: Collagen was rehydrated with approximately 100 µL 50 mM ammonium bicarbonate per milligram and each digested with 0.4 µg sequencing grade trypsin (Promega,

289

290

291

292

293

294

295

296

297

298

299

300

301

302

303

304

305

306

307

308

309

310

311

UK) overnight at 37°C. Initial attempts were made at directly spotting these digests through

dilution in 0.1% trifluoroacetic acid (TFA; Buckley, et al. 2018), but many required ZipTip purification with C18 solid phase extraction and rehydration in 0.1% TFA (Buckley, et al. 2009). Samples were spotted onto a stainless-steel Matrix Assisted Laser Desorption Ionization (MALDI) target plate with equal volume of 10 mg/mL alpha-cyano hydroxycinnamic acid in 50% acetonitrile/0.1% TFA and allowed to air dry. Dried spots were then analyzed using a Bruker Ultraflex II MALDI mass spectrometer over the m/z range 700-3700 with up to 2,000 laser acquisitions for each sample. Peptide mass fingerprints were then compared with those of standard reference material via the identification of genus- or species-specific biomarkers (SI 4). Statistical analyses: For statistical comparisons of fish  $\delta^{15}N$  values, each taxon (i.e., lake trout, Atlantic salmon, whitefishes) was compared separately by time periods binned by 100 (pre-1925 CE) and 25 (post-1925 to present) year groups (SI 5, Table S1) using PAST version 3.22. Bin timeframes were selected with a view to maximizing sample size per bin. To assess whether fish δ<sup>15</sup>N values changed over time we first compared pre-1800 bins to one another to establish whether significant differences occur within each fish taxon between sequential time frames. After finding that no significant differences occur (SI 5, Table S2), we then grouped all pre-1775 (preindustrial) bins and compared this individually to all succeeding time periods (SI 5, Table S3). For each bin, normality of distribution was first tested using a Shapiro-Wilk's test (SI 5, Table S1). For datasets where all time periods were distributed normally, a one-way ANOVA was used, and homogeneity of variance was assessed using a Levene's test. A post hoc Dunnett's T3 test or a post hoc Tukey's HSD test was used to compare groups with variances that were equal or unequal, respectively. For data sets where not all time periods were distributed normally, Mann-Whitney U tests with Bonferroni corrected p values were used to compare groups.

312

313

314

315

316

317

318

319

320

321

322

323

324

325

326

327

328

329

330

331

332

333

#### Results

335

336

337

338

339

340

341

342

343

344

345

346

347

348

349

350

351

352

353

354

355

356

357

Of the samples with sufficient collagen (n=607 of 662) for isotopic analyses, 96% (total n=582) produced acceptable C:N, %C, and %N values satisfying quality control criteria. Interpretations also include previously published data from archaeological (n=68; (Guiry, et al. 2016a; van der Merwe, et al. 2003)) and modern (post-1958 CE; n=819; (Colborne, et al. 2016; Fera, et al. 2017; Kiriluk, et al. 1995; Lumb and Johnson 2012; Mumby, et al. 2018; Rush, et al. 2012; Yuille, et al. 2015)) specimens. ZooMS analyses of 290 samples confirmed or helped to refine 244 nonspecies taxonomic identifications provided through bone morphology analyses zooarchaeologists (it confirmed the identification for 225 of these 244 specimens and refined it to the level of species for the remaining 19). In a further 20 cases, specimens were reassigned to another taxon, namely, 12 to lake trout (Salvelinus namaycush) (previously assigned with varying degrees of confidence to the species or genus level in Salmo [n=8], Salvelinus [n=2], or Coregonus [n=2]); four to Atlantic salmon (Salmo salar) (previously identified with varying degrees of confidence as Salvelinus namaycush); four to whitefish (C. clupeaformis or C. artedi) (previously assigned with varying degrees of confidence to the species or genus level in Salmo salar [n = 1], Salvelinus fontinalis [n = 2], or Salvelinus namaycush [n = 1]). In 15 other cases, ZooMS could not reassign specimens (previously assigned with varying degrees of confidence to the Salmonidae family level [n = 3] or to the species or genus level in Salmo [n=4], Salvelinus [n=1], or Coregonus [n=4]) to one of the four species and one genus used in this study. Based on variation in peptide mass fingerprints, these samples likely derive from at least eleven other species. Stable nitrogen isotope compositions of bone and scale collagen from archaeological and

historical lake trout (n=196), Atlantic salmon (n=147), and whitefishes (n=239) are summarized

in the SI 5, Table S1 and presented in full in Figure 2 and SI 2. Given the large and well-dated sample (n=1469) from archaeological sites (n=42) and from museum-archived specimens and contemporary fisheries research (n=<50 locations) across a broad geographical range, we believe that the temporal trends observed in fish bone and scale collagen  $\delta^{15}N$  are representative of processes occurring in Lake Ontario's open water ecosystem at a regional scale.

#### Discussion and conclusion

358

359

360

361

362

363

364

365

366

367

368

369

370

371

372

373

374

375

376

377

378

379

380

1200–1830 CE: Fish  $\delta^{15}$ N shows remarkably little variation between 1200 CE and 1830 CE (Figure 2), despite centuries of Indigenous and European land management in the Lake Ontario watershed, and provides a unique perspective on the long-term stability of a Great Lakes ecosystem. Prior to the nineteenth century, mean  $\delta^{15}N$  values binned by taxon and at 100-year intervals vary by less than 0.4% and show no statistically significant differences (SI 5, Tables S1 and S2). This long-term stasis, over at least 600 years, suggests that the nitrogen cycle and nitrogenous nutrient inputs for Lake Ontario were highly stable during the past millennium. Analyses of two additional Atlantic salmon specimens (not shown in Figure 2) from earlier archaeological deposits suggests that this stability extends further back in time to at least 900 CE. However, due to a paucity of samples from the earlier time period, discussion will focus on the post-1200 CE time frame. A comparison of mean  $\delta^{15}$ N for pre-1800 bins (n=4 see SI 5, Table S1) shows steady offsets between species: lake trout  $+1.6\pm0.2\%$  > Atlantic salmon  $+1.3\pm0.2\%$  > whitefishes. Replication of this pattern across all three taxa, encompassing multiple trophic levels, suggests that stability was also characteristic of interspecific trophic relationships throughout the broader pelagic food web during this period.

A wealth of historical and paleoenvironmental research on the Lake Ontario watershed (Beeton

1965; Bogue 2001; Estepp and Reavie 2015; Hodell and Schelske 1998; Jeffers, et al. 2015;

Meyers 2003; Schelske and Hodell 1991b; Schelske and Hodell 1995; Schelske, et al. 1988; Schelske, et al. 1983; Wood 2000) provides an excellent opportunity to contextualize and consider the implications of this long-term stability. Proxies for past environmental conditions in other, smaller lakes in the same region document anthropogenic disturbances, at least as early as the thirteenth century CE, resulting from Indigenous land clearance and farming activities (e.g., Ekdahl, et al. 2004). These studies suggest that Indigenous land management practices, including the use of fire for forest clearance followed by crop cultivation (Munoz and Gajewski 2010), had a significant impact on soil erosion, sedimentation, and nutrient loading, causing early cultural eutrophication of aquatic environments in the region. European settlement of the Lake Ontario watershed began in the seventeenth century (Gentilcore 1984). Settlement expansion in the nineteenth century was accompanied by intensive commercial fishing (1800 onward; Bogue 2001), diversion of the Niagara River and work on the Erie and Welland canals (1820s; McIlwraith 1976; Wood 2000), broad-scale land clearance for lumber and agriculture (1840s-1850s; Head 1975; Lower, et al. 1938), and wetland loss due to mining and other activities (1880s; Bogue 2001). Paleolimnological studies suggest that productivity of Lake Ontario's phytoplankton community began to grow slowly with the first European settlement, with more substantial impacts occurring in the mid-nineteenth century, probably in response to nutrient loading from soil erosion following deforestation (e.g., Estepp and Reavie 2015; Hodell, et al. 1998; Schelske 1991; SI 1). Despite clear historical and paleolimnological evidence for earlier anthropogenic impacts on nutrient dynamics (e.g., nitrogen and phosphorus loading, pollen species change) in other areas of the watershed (Bunting, et al. 1998; Duthie and Sreenivasa 1971; Ekdahl, et al. 2004; Ekdahl, et al. 2007; Munoz and Gajewski 2010; Schelske, et al. 1983; Yang, et al. 1993), archaeological fish stable nitrogen isotopic compositions demonstrate that

381

382

383

384

385

386

387

388

389

390

391

392

393

394

395

396

397

398

399

400

401

402

Lake Ontario's nitrogen cycle and food web structure remained unchanged until the 1830s. In that context, and given the sensitivity of aquatic nitrogen cycles to human disturbances (Anderson and Cabana 2005; Botrel, et al. 2014; Lake, et al. 2001), these data underscore Lake Ontario's stability throughout much of the last millennium and demonstrate a degree of resilience at the level of the nitrogen cycle in large lake ecosystems to a variety of human impacts, even when sustained over long time spans.

The resilience of Lake Ontario's nitrogen cycle has important implications for understanding

404

405

406

407

408

409

410

411

412

413

414

415

416

417

418

419

420

421

422

423

424

425

426

human impacts on freshwater ecosystems in the recent and more ancient past. It is now well known that industrial activities over the past 150 years have fundamentally altered global nutrient distributions and biogeochemical cycles (Galloway and Cowling 2002). Within the context of debate about the onset of the Anthropocene (Lewis and Maslin 2015), there is a growing consensus that "pristine environments" probably did not exist where humans were present, because there was always some form of impact from human activities (Heckenberger, et al. 2003). This perspective has been supported by archaeological research showing that ancient societies, particularly those that used agriculture, had significant impacts at both local and regional scales on terrestrial and aquatic nutrient cycles for millennia (Curtis, et al. 1998; Guiry, et al. 2018; Hadley, et al. 2010). Evaluating the role of past human activities as drivers of ecological change has been a priority for archaeologists (Kintigh, et al. 2014), but is complicated by the fact that ancient anthropogenic impacts on the environment are spatiotemporally heterogeneous and occur along a continuum of intensity (from very significant to ephemeral). For ecologists, this issue represents a source of uncertainty for retrospective studies where archaeological or paleobiological information is not available to establish when and to what extent past human activities have impacted a particular environment or biotic community

(Holmes 2006; Lyman and Cannon 2004). In contrast to the cautionary tone of the debate about 427 the existence of "pristine" environments, our data demonstrate that, in fact, nutrient dynamics 428 and trophic relationships in larger aquatic environments can remain unchanged despite centuries 429 of moderate human land use. 430 Post-1830: Our analyses of archaeological and historical fish also show that this long-term 431 stability was profoundly altered when Lake Ontario's nutrient pools experienced an abrupt shift 432 in stable nitrogen isotope composition, becoming enriched in <sup>15</sup>N throughout the pelagic 433 ecosystem. During the early to mid-nineteenth-century, fish  $\delta^{15}N$  values underwent a 434 simultaneous, statistically significant elevation across multiple trophic levels, a difference that 435 has been sustained throughout the twentieth century and to this day (SI 4, Table S3). Atlantic 436 salmon, for instance, which had previously had a highly conserved distribution of  $\delta^{15}$ N values, 437 show an elevation of +3\% in well-dated mid-nineteenth-century museum specimens. Although a 438 paucity of fish samples from the later nineteenth and early to mid-twentieth century precludes 439 detailed comparison, these data show similar overall trends (positive shifts between intraspecific 440  $\delta^{15}$ N means for pre-1800 and post-1990, ranging from +5.3% to +6.2%, for all fish species) to those observed in Lake Ontario's isotopic records from sediment cores (+5.5% to +5.6% for c. 442 1850 to 1990s; Hodell and Schelske 1998; Figure 2). This further indicates that the observed 443 upward pattern in fish  $\delta^{15}N$  predominately reflects a bottom-up shift in baseline  $\delta^{15}N$  rather than 444 changes to fish trophic behavior. 445 446 It is nonetheless apparent from these data that human activities have resulted in meaningful behavioral changes, including increased niche overlap between Lake Ontario's salmonid 447 communities. Prior to 1800, there is minimal overlap between the  $\delta^{15}N$  ranges for each species, 448

441

449

with overlaps of only 0.1% between Atlantic salmon and whitefishes and only 0.9% between

Atlantic salmon and lake trout over a 600-year period. In contrast, in the twentieth century, there is substantial overlap between  $\delta^{15}N$  ranges for all species in all 25-year temporal bins (overlap ranges from 1.7 to 4.7%). However, caution is required when interpreting trends in variation across time in this dataset. Differences in tissue turnover rates for sample materials between prenineteenth- (mainly collagen from bones - slow turnover) and twentieth-century (mainly previously published muscle from modern fish - fast turnover) time frames make detailed analysis of these trends difficult because tissues with faster turnover rates are inherently more likely to produce greater isotopic variation reflecting seasonal or other short-term dietary aberrations. With that caveat in mind, we argue that it is likely that the greater isotopic variation in twentieth-century specimens reflects changes in energy pathways and foraging behavior in response to major anthropogenic environmental disruptions (e.g., impacts from invasive species, extirpations, or over fishing; Bogue 2001; Colborne, et al. 2016; Dymond, et al. 2019; Fera, et al. 2017; Mills, et al. 2003). The interspecific synchronicity and magnitude of this shift could have been caused by multiple related processes (Figure 3), including changes in the nitrogen cycle (at the phytoplankton-DIN pool level) favoring nitrogen transformations that retain <sup>15</sup>N and changes in nitrogen inputs to the system that include <sup>15</sup>N-enriched nutrients (Finlay and Kendall 2007; Kendall 1998; Kendall, et al. 2007; Talbot 2001). Detailed historical and archaeological dating provides a robust temporal framework to evaluate which of these variables, and therefore what type of human activity, drove this initial ecosystem-wide <sup>15</sup>N enrichment in Lake Ontario's food web. At archaeological sites with occupation dates as late as 1832, many fish show no sign of  $\delta^{15}N$  elevation for their respective taxon, whereas a museum specimen dating from no later than 1857 shows significantly elevated  $\delta^{15}$ N relative to the preceding centuries. In addition, many specimens from

450

451

452

453

454

455

456

457

458

459

460

461

462

463

464

465

466

467

468

469

470

471

archaeological contexts with date ranges spanning this terminus ante quem (1832) and terminus post quem (1857) show elevated  $\delta^{15}$ N values. These data therefore provide a temporal bracket for the initiation of change in Lake Ontario's nitrogen cycle, demonstrating that the shift occurred abruptly (within the context of the stability of the preceding 900 years), within a relatively short, 15–25-year window (1830s–1850s). This timeframe coincides precisely with the emergence of broad-scale land clearance for agriculture and timber harvesting in the Lake Ontario watershed. From the 1830s onward, especially around the middle of the century, historical analyses document how forestry activities, namely severe cutting as well as widespread burning of the remaining slash, radiated outward from the Lake Ontario waterfront to encompass much of the watershed. These forestry activities accelerated over time due to increased demand for timber (Head 1975) and cleared land, improved export capacity (canals and railroad; Lower, et al. 1938; McIlwraith 1976), and growing sawmill infrastructure (Head 1975; Wood 2000). During this period, between the 1820s and 1850s, the then-province of Upper Canada (which included the Ontario portion of the Lake Ontario watershed) had the fastest growing population in all of North America (Lewis 2001). Prior to 1830, newly settled farmers of European heritage focused on subsistence (Kelly 1973; Kelly 1975) and put considerable effort into clearing land for cultivation through cutting and burning (Ball 1979). However, relative to the mid-nineteenth century, these early farming activities were patchy, small in scale, and diversified in purpose, with most of the land granted to settlers remaining forested (Kelly 1975). Moreover, much of the land that was cleared retained stabilizing features of the previous forest, including stumps, root mats, and stones, for up to a decade, and sometimes much longer, while under cultivation and these would have mitigated issues with soil erosion and nutrient loss (Ball 1979; Kelly 1975). During this early period, in which the farmers were themselves clearing their land, care was

473

474

475

476

477

478

479

480

481

482

483

484

485

486

487

488

489

490

491

492

493

494

often taken throughout the processes, from cutting trees to sowing crops (delaying ploughing), to further minimize soil nutrient loss (Ball 1979; Kelly 1971). As farms became established, however, stumps were removed, ploughs were more widely employed to break up the soil, and more land came under cultivation. Throughout the 1830s and 1840s, the agricultural system of Upper Canada was dominated by plough-intensive (three or more times per biennial fallow) and highly profitable monocropping of wheat (Kelly 1971; Kelly 1973). The focus on a wheat-fallow-wheat agriculture system, as well as labor-saving furrow drainage and general avoidance of manure-based crop fertilization (until the 1850s) were widely blamed for rapid depletion of soil nutrients as well as erosion across the region (Kelly 1971; Kelly 1975). In this context, prior to the 1880s, when a wider appreciation developed among farmers in the Lake Ontario watershed of the risks of higher runoff-to-percolation ratios that come with overly cleared lands, farmland was subjected to a damaging seasonal cycle of spring floods, washing away nutrient-rich organic materials, and summer droughts, parching what soil remained (Kelly 1975).

The isotopic composition of reactive forms of nitrogen in terrestrial ecosystems (i.e., soil nutrients including ammonia and nitrate), particularly those used for agriculture, is typically elevated relative to aquatic nitrogen sources (i.e., DIN) due to intense soil denitrification (Botrel, et al. 2014; Heaton 1986; Kendall 1998). In this context, the most parsimonious explanation for a broad-scale isotopic shift in Lake Ontario's nitrogen cycle is that there was an increase in <sup>15</sup>N-enriched nutrient inputs coming directly from soil erosion following deforestation during the 1840s in Lake Ontario's watershed area (Figure 3). This interpretation is supported by paleolimnological data (diatoms, sediment accumulation rates, and elemental and isotopic compositions of organic matter) from across the Lake Ontario watershed that records an uptick in nutrient loading from soil erosion runoff over the course of the mid-nineteenth century (Hodell

and Schelske 1998). The relatively early timeframe for the observed shift in fish nitrogen isotopic compositions also suggests that its cause lay in new nutrient inputs originating primarily within the local watershed area, rather than from other Great Lakes, because upstream watersheds such as Lake Erie had not been as extensively settled by this time.

519

520

521

522

523

524

525

526

527

528

529

530

531

532

533

534

535

536

537

538

539

540

While inputs of <sup>15</sup>N-enriched terrestrial nutrients, released from eroding soils across the watershed, could easily result in the observed isotopic pattern, additional related processes, occurring both on land and in the water, could further contribute to the elevation of the stable nitrogen isotopic composition of Lake Ontario's biota (Figure 3). Higher soil erosion rates would increase mobilization of terrestrial nitrate across oxygen-poor ecotones (i.e., riparian/wetland zones), promoting bacterial denitrification, leaving the residual nutrients that are eventually delivered to Lake Ontario relatively enriched in <sup>15</sup>N (Bowden 1987; Knowles 1982; McClain, et al. 2003). Increases in anthropogenic soil disturbance from farming and use of fertilizers (pushing the nitrogen cycle to be more open; Szpak 2014), as well as changes to pH in aquatic environments, may promote volatilization of ammonia before or during nutrient transport to Lake Ontario's food web (Kendall 1998). Increased phosphorous loading from soil runoff would also increase primary productivity, and the resulting bottom-up shift would have increased demand on Lake Ontario's pool of available reactive N, thereby reducing discrimination against <sup>15</sup>N during assimilation of DIN by phytoplankton (Hodell and Schelske 1998). Regardless of which of these processes were most facilitative of the nitrogen isotopic patterning in Lake Ontario's fish community, the timing (between 1832 and 1857) and abruptness of the ecosystemwide shift indicates that the first human activity to have broad-scale impacts on nitrogenous nutrient pathways in the Great Lakes was likely deforestation.

It is also worth noting that the time period covered by this study includes significant global climatic fluctuations, including the Medieval Climate Anomaly (MCA; 950-1250 CE) and the Little Ice Age (LIA; 1300–1850 CE). Paleolimnological studies of Lake Ontario (McFadden, et al. 2005), Lake Erie (Finkelstein and Davis 2006), and other lakes in the local (Keizer, et al. 2015; Mullins, et al. 2011; Paquette and Gajewski 2013; Stager, et al. 2017) and broader region (Laird, et al. 2012) show that climatic variability associated with both the MCA and LIA did have some impact on aquatic and terrestrial environments across the Great Lakes and St. Lawrence River region. For instance, evidence from flux in biogenic silica sedimentation rates and changes in diatom (e.g., Fragelaria spp.) communities (Finkelstein, et al. 2005; McFadden, et al. 2005) demonstrates that these periods of climatic change were felt in the Lake Ontario watershed over the past 1000 years. Climatic variability could affect Lake Ontario's nitrogen cycle and the nitrogen isotopic composition of fish through changes in adjacent wetland size (important sites for denitrification; Bowden 1987; McClain, et al. 2003), terrestrial runoff impacts (Anderson and Cabana 2005; Heaton 1986), fluctuating dissolved oxygen levels (controlling nitrification and denitrification rates; Knowles 1982), and changes to productivity and thermally regulated stratification (seasonal draw-down of DIN; Hodell and Schelske 1998). Assuming that the influence of these potential sources of environmental variation did not cancel one another out, our data suggest that late Holocene climatic variation did not influence Lake Ontario's nitrogen cycle or trophic structure. The temporal as well as taxonomical detail offered by isotopic data from archaeological fish specimens has provided a framework for higher resolution interpretation of food web dynamics as well as the timing and likely cause of the first human impacts on Lake Ontario's nitrogen cycle. While previous isotopic analyses of organic matter from sediments (SI 1 and Figure 2;

541

542

543

544

545

546

547

548

549

550

551

552

553

554

555

556

557

558

559

560

561

562

Hodell and Schelske 1998; McFadden, et al. 2004) have demonstrated that Lake Ontario's nitrogenous nutrients were becoming <sup>15</sup>N enriched during the nineteenth century, temporal differences between datasets as well as a dearth of pre-1850 data prevented detailed analyses of the processes which drove this change in lake-wide nitrogen cycling (SI 1). In addition, although a variety of paleoenvironmental indicators (e.g., biogenic silica; Schelske 1991; Schelske and Hodell 1991b; Schelske, et al. 1988) have shown human activities, such as deforestation, had begun to have major nutrient (particularly P) loading impacts on the Lake Ontario watershed from the mid-nineteenth century onward (although see, McAndrews and Boyko 1972; Schelske, et al. 1983), our results demonstrate that Lake Ontario's nitrogen cycle had already been substantially impacted decades earlier. Finally, by analyzing material from known taxa, we have been able to document the impact that these changes in lake-wide nutrient dynamics had on longterm food web structure. Anthropological and Ecological Implications: This study is the first detailed analysis of longterm isotopic variation in freshwater fish. Results suggest that, by altering aquatic nutrient inputs, nineteenth-century European forestry practices in the lower Great Lakes region, whether focused on timber extraction or producing cleared land for agriculture, left a durable isotopic signature on fish in Lake Ontario. While the findings of this research focus on the Great Lakes watershed, the implications are global; human impacts on aquatic nutrient dynamics, particularly the nitrogen cycle, may be detectable in archaeological fish remains wherever land management has extensively modified the balance of nutrient exchange between local terrestrial and aquatic landscapes. Our results are particularly germane to recent discussion in conservation, calling for increased attention to how human impacts on land may be a driver of environmental

564

565

566

567

568

569

570

571

572

573

574

575

576

577

578

579

580

581

582

583

584

585

586

deterioration in adjacent aquatic ecosystems (Abell 2002). For this reason, our findings have

significant potential to serve as a model for research that seeks to understand fundamental changes in the way past societies, through forestry and other land management practices, affected aquatic nutrient dynamics around the globe (Jenny, et al. 2019).

With respect to debate on the origins of the Anthropocene, our results provide an important waypoint for the earliest lasting anthropogenic alteration of one of the world's great freshwater ecosystems. By identifying the tipping point at which human activities began to reorganize nutrient dynamics in the Great Lakes region, our results do not reveal a "golden spike" indictor for the starting point of the Anthropocene (Lewis and Maslin 2015), but, rather, provide a new signpost along the path of the global expansion of industrial impacts. These changes to Lake Ontario nonetheless represent a profound shift in the way humans have impacted the Great Lakes environment and illustrate some important anthropological and ecological considerations for how we can approach aquatic human—environment relationships in archaeology and historical ecology moving forward.

From an ecological perspective, an ecosystem-wide, bottom-up alteration of the nitrogen cycle of one of the world's largest freshwater environments over the course of only one or two decades, as shown here for Lake Ontario, demonstrates an unprecedented shift in the nature and scale of how humans cause environmental change. Understanding how humans have shaped earth's biological and physical systems remains an important archaeological and ecological challenge because finding lasting evidence of human impacts within complex regional or global systems is rare. This is particularly true for our understanding of ancient environmental nutrient dynamics, such as the cycling of nitrogen throughout an ecosystem, which, although fundamental to the success of human societies both today and in the past, preserves relatively few traces of change. Not only do our findings demonstrate how isotopic analyses of archaeological fauna can provide

a clearer overview of ancient nutrient dynamics, they do so in one of the most enigmatic arenas of human-environment interaction, namely, aquatic ecosystems. Even today, in comparison with terrestrial environments, surprisingly little is known about how escalating human exploitation of the world's aquatic environments will impact these vital ecosystems in the long term (Abell 2002). In that context, deeper retrospectives that are based on integrating results of contemporary studies with data from both historical and archaeological resources, as demonstrated by this study, have the potential to significantly enhance our understanding of the sensitivity of different environments to human disturbances (Barak, et al. 2016; Dubois, et al. 2018; Jeffers, et al. 2015; McLauchlan, et al. 2013). From an anthropological perspective, approaching questions of human–environment interactions from the vantage point of nutrient dynamics could provide a novel outlook on the role of humans as drivers of ecological change. While the relationship between humans and their environment has traditionally been framed against a backdrop of such factors as technological change and the development of social complexity as sources of evidence for macro-scale exploitation of environmental resources (Kintigh, et al. 2014), there is an increasing recognition that an understanding of how humans have influenced their landscape at a molecular level can provide powerful new lines of evidence for previously invisible but profound shifts in humanenvironment relations (Guiry, et al. 2018). In particular, throughout most times and places in human history, the carrying capacity of local terrestrial and aquatic ecosystems, which are the environmental foundations upon which societies are sustained, has hinged on the maintenance of well-balanced biogeochemical cycles and nutrient regimes. With respect to lake and other freshwater ecosystems, which have been and continue to be of pivotal importance to many cultures, it is possible to use isotopic analyses of local fauna to look back in time and pinpoint

610

611

612

613

614

615

616

617

618

619

620

621

622

623

624

625

626

627

628

629

630

631

- where, when, and in that context perhaps even how, human activities reached a turning point to
- become dominant drivers in their ecosystem.

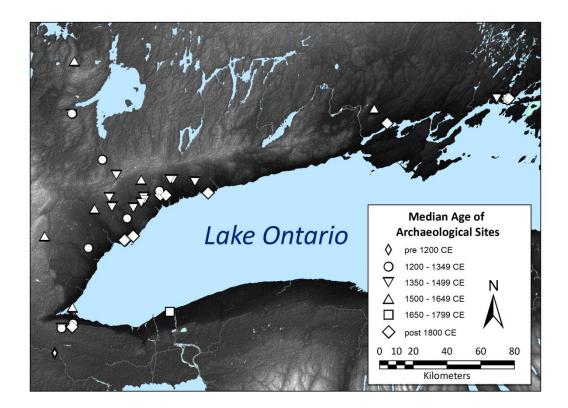
636

633

#### Acknowledgments

Sampling permissions and logistical assistance: This analysis entailed destructive analysis of 637 zooarchaeological specimens from Huron-Wendat and St. Lawrence Iroquoian sites. We thank 638 representatives of descendant First Nations (Louis Lesage, Huron-Wendat Nation and Henry 639 Lickers, Mohawk Council of Akwesasne) for permission for this analysis. We also thank Hernan 640 Lopez-Fernandez and the Royal Ontario Museum; Shari Prowse and the Ministry of Tourism, 641 Culture and Sport; Ron Williamson, Alexis Dunlop, Caitlin Coleman and Archaeological 642 Services Inc.; Dena Doroszenko and the Ontario Heritage Trust; Shaun Austin and Wood 643 Environment & Infrastructure Solutions; Stacey Girling-Christie, Jean-Luc Pilon and the 644 Canadian Museum of History; Jennifer Campbell, Meaghan Eckersley and the City of Kingston; 645 Lena Beliveau, Ashley Mendes and the Royal Military College; Loren Scott, Cassandra 646 Hamilton, Janet Batchelor and the Toronto Region Conservation Authority; David G. Smith, 647 Heather M.-L. Miller and the Department of Anthropology, University of Toronto Mississauga; 648 Andrew Martindale and Jessica Metcalf, University of British Columbia. Funding: Department 649 of Anthropology, University of British Columbia; Social Sciences and Humanities Research 650 Council of Canada (SSHRC) Insight Development Grant. EG was supported by a SSHRC 651 Postdoctoral Research Fellowship, a SSHRC Banting Postdoctoral Research Fellowship, and the 652 653 Department of Anthropology at the University of British Columbia. MB was supported by a Royal Society University Research Fellowship. Author Contributions: E.G. designed research. 654 S.N.-H., A.H., T.O., E.H., and E.G. contributed samples for analysis. E.G., M.B., P.S. performed 655 isotopic, ZooMS, and statistical analyses, respectively. E.G. interpreted results and wrote the 656 paper with assistance from all authors. Figures were created by A.H. (1) and E.G. (2 and 3). 657 Competing Interests: The authors declare no competing interests. Data Accessibility: All data 658 generated as part of this research is available in the SI. We thank two anonymous reviewers for 659 their helpful comments. 660

## Figure Captions



**Figure 1. Map of study area** showing locations of archaeological sites from which fish bone samples were collected. Map data from NASA and ESRI.

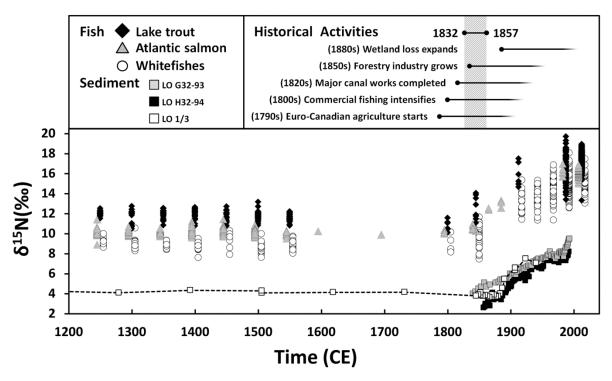
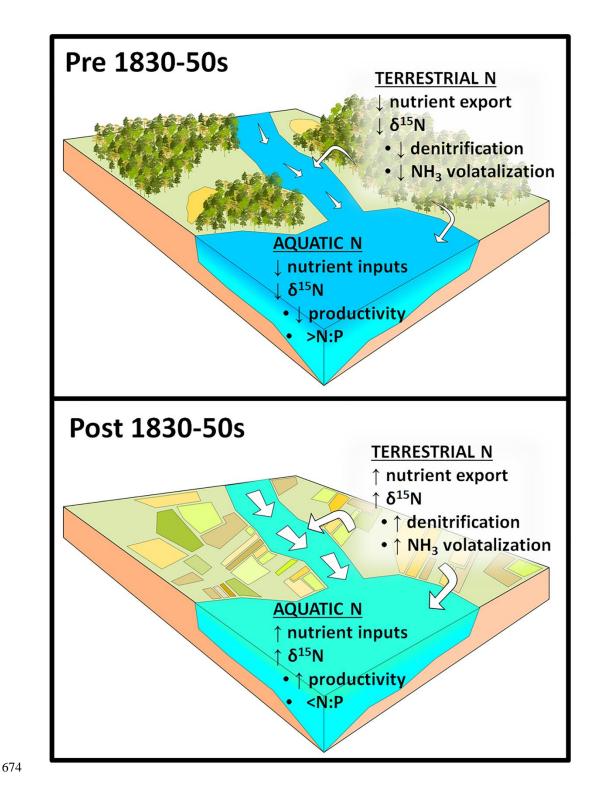


Figure 2. Stable nitrogen isotope values for fish (*n*=1469) versus time. Data are binned by 50-year (archaeological) and 25-year (historical/modern) intervals. For fish data sourced from the literature (Colborne, et al. 2016; Fera, et al. 2017; Guiry, et al. 2016a; Kiriluk, et al. 1995; Lumb and Johnson 2012; Mumby, et al. 2018; Rush, et al. 2012; van der Merwe, et al. 2003; Yuille, et al. 2015). For sediment data sourced from the literature: LO G32-93, LO H32-94 (Hodell and Schelske 1998), and LO 1/3 (McFadden, et al. 2004).



**Figure 3. Schematic diagram of nutrient regime shifts** (nitrogen sources and isotopic compositions) that could be associated with the intensification of deforestation and agriculture in the Lake Ontario watershed during the early to mid-nineteenth century.

676

678	Supplementary Information
679	SI1 Paleoenvironmental context
680	SI2 Isotopic results
681	SI3 Calibration and analytical uncertainty
682	SI4 Collagen peptide finger-printing results
683	SI5 Statistical results
684	
685	References
686	Abell R. 2002. Conservation biology for the biodiversity crisis: a freshwater follow-up.
687	Conservation Biology 16(5):1435-1437.
688	Allan JD, McIntyre PB, Smith SDP, Halpern BS, Boyer GL, Buchsbaum A, Burton GA,
689	Campbell LM, Chadderton WL, Ciborowski JJH and others. 2013. Joint analysis of
690	Stressors and ecosystem services to enhance restoration effectiveness. Proceedings of the
691	National Academy of Sciences of the United States of America 110(1):372-377.
692	Anas MUM, Simpson GL, Leavitt PR, Cumming BF, Laird KR, Scott KA, Das B, Wolfe JD,
693	Hesjedal B, Mushet GR and others. 2019. Taxon-specific variation in δ13C and δ15N of
694	subfossil invertebrate remains: Insights into historical trophodynamics in lake food-webs.
695	Ecological Indicators 102:834-847.
696	Anderson C, Cabana G. 2005. δ15N in riverine food webs: effects of N inputs from agricultural
697	watersheds. Canadian Journal of Fisheries and Aquatic Sciences 62(2):333-340.
698	Anderson NJ. 2014. Landscape disturbance and lake response: temporal and spatial perspectives.
699	Freshwater Reviews 7(2):77-120.
700	Ball NR. 1979. The technology of settlement and land clearing in Upper Canada prior to 1840:
701	University of Toronto.  Barak RS, Hipp AL, Cavender-Bares J, Pearse WD, Hotchkiss SC, Lynch EA, Callaway JC,
702 703	Calcote R, Larkin DJ. 2016. Taking the long view: integrating recorded, archeological,
703 704	paleoecological, and evolutionary data into ecological restoration. International Journal
704	of Plant Sciences 177(1):90-102.
705	Beeton AM. 1965. Eutrophication of the St. Lawrence great lakes. Limnology and Oceanography
707	10(2):240-254.
707	Bogue MB. 2001. Fishing the Great Lakes: an environmental history, 1783–1933: Univ of
709	Wisconsin Press.
710	Botrel M, Gregory-Eaves I, Maranger R. 2014. Defining drivers of nitrogen stable isotopes
711	(δ15N) of surface sediments in temperate lakes. Journal of Paleolimnology 52(4):419-
712	433.
713	Bowden WB. 1987. The biogeochemistry of nitrogen in freshwater wetlands. Biogeochemistry
714	4(3):313-348.

- Buckley M, Collins M, Thomas- Oates J, Wilson JC. 2009. Species identification by analysis of bone collagen using matrix- assisted laser desorption/ionisation time- of- flight mass spectrometry. Rapid communications in mass spectrometry 23(23):3843-3854.
- Buckley M, Gu M, Herman J, Junno J-A, Denys C, Chamberlain AT. 2018. Species identification of voles and lemmings from Late Pleistocene deposits in Pin Hole Cave (Creswell Crags, UK) using collagen fingerprinting. Quaternary international 483:83-89.

- Bump JK, Fox-Dobbs K, Bada JL, Koch PL, Peterson RO, Vucetich JA. 2007. Stable isotopes, ecological integration and environmental change: wolves record atmospheric carbon isotope trend better than tree rings. Proceedings of the Royal Society of London B: Biological Sciences 274(1624):2471-2480.
- Bunting MJ, Morgan CR, Bakel MV, Warner BG. 1998. Pre-European settlement conditions and human disturbance of a coniferous swamp in southern Ontario. Canadian Journal of Botany 76(10):1770-1779.
- Canfield DE, Glazer AN, Falkowski PG. 2010. The evolution and future of Earth's nitrogen cycle. science 330(6001):192-196.
- Colborne SF, Rush SA, Paterson G, Johnson TB, Lantry BF, Fisk AT. 2016. Estimates of lake trout (Salvelinus namaycush) diet in Lake Ontario using two and three isotope mixing models. Journal of Great Lakes Research 42(3):695-702.
- Collister JW, Hayes J. 1991. A preliminary study of the carbon and nitrogen isotopic biogeochemistry of lacustrine sedimentary rocks from the Green River Formation, Wyoming, Utah, and Colorado.
- Curtis JH, Brenner M, Hodell DA, Balser RA, Islebe GA, Hooghiemstra H. 1998. A multi-proxy study of Holocene environmental change in the Maya Lowlands of Peten, Guatemala. Journal of paleolimnology 19(2):139-159.
- DeNiro MJ. 1985. Postmortem preservation and alteration of in vivo bone collagen isotope ratios in relation to palaeodietary reconstruction. Nature 317:806-809.
- Denk TR, Mohn J, Decock C, Lewicka-Szczebak D, Harris E, Butterbach-Bahl K, Kiese R, Wolf B. 2017. The nitrogen cycle: A review of isotope effects and isotope modeling approaches. Soil Biology and Biochemistry 105:121-137.
- Dubois N, Saulnier-Talbot É, Mills K, Gell P, Battarbee R, Bennion H, Chawchai S, Dong X, Francus P, Flower R. 2018. First human impacts and responses of aquatic systems: A review of palaeolimnological records from around the world. The Anthropocene Review 5(1):28-68.
- Duthie HC, Sreenivasa M. 1971. Evidence for the eutrophication of Lake Ontario from the sedimentary diatom succession. In: Duthie HC, Yang, J.R., Edwards, editor. Proceedings, 14th Conference on Great Lakes Research. Ann Arbor, Michigan: International Association for Great Lakes Research. p 1–13.
- Dymond JR, MacKay HH, Burridge ME, Holm E, Bird PW. 2019. The history of the Atlantic Salmon in Lake Ontario. Aquatic Ecosystem Health & Management(just-accepted):1-12.
- Ekdahl EJ, Teranes JL, Guilderson TP, Turton CL, McAndrews JH, Wittkop CA, Stoermer EF. 2004. Prehistorical record of cultural eutrophication from Crawford Lake, Canada. Geology 32(9):745-748.
- Ekdahl EJ, Teranes JL, Wittkop CA, Stoermer EF, Reavie ED, Smol JP. 2007. Diatom assemblage response to Iroquoian and Euro-Canadian eutrophication of Crawford Lake, Ontario, Canada. Journal of Paleolimnology 37(2):233-246.

Estepp LR, Reavie ED. 2015. The ecological history of Lake Ontario according to phytoplankton. Journal of Great Lakes Research 41(3):669-687.

767

768

769

777

778

779

780

783

784 785

786

787

788

789

790

791 792

793

794

795

796

797

798

- Fera SA, Rennie MD, Dunlop ES. 2017. Broad shifts in the resource use of a commercially harvested fish following the invasion of dreissenid mussels. Ecology 98(6):1681-1692.
- Finkelstein SA, Davis AM. 2006. Paleoenvironmental records of water level and climatic changes from the middle to late Holocene at a Lake Erie coastal wetland, Ontario, Canada. Quaternary Research 65(1):33-43.
  - Finkelstein SA, Peros MC, Davis AM. 2005. Late Holocene paleoenvironmental change in a Great Lakes coastal wetland: integrating pollen and diatom datasets. Journal of Paleolimnology 33(1):1-12.
- Finlay JC, Kendall C. 2007. Stable isotope tracing of temporal and spatial variability in organic matter sources to freshwater ecosystems. Stable isotopes in ecology and environmental science 2:283-333.
- Frossard V, Verneaux V, Millet L, Jenny JP, Arnaud F, Magny M, Perga ME. 2014.
  Reconstructing long- term changes (150 years) in the carbon cycle of a clear- water lake based on the stable carbon isotope composition (δ13C) of chironomid and cladoceran subfossil remains. Freshwater biology 59(4):789-802.
  - Galloway JN, Cowling EB. 2002. Reactive nitrogen and the world: 200 years of change. AMBIO: A Journal of the Human Environment 31(2):64-71.
  - Gentilcore RL. 1984. The making of a province: Ontario to 1850. American Review of Canadian Studies 14(2):137-156.
- Gruber N, Galloway JN. 2008. An Earth-system perspective of the global nitrogen cycle. Nature 451(7176):293.
  - Gu B. 2009. Variations and controls of nitrogen stable isotopes in particulate organic matter of lakes. Oecologia 160(3):421-431.
  - Guiry E. 2019. Complexities of Stable Carbon and Nitrogen Isotope Biogeochemistry in Ancient Freshwater Ecosystems: Implications for the Study of Past Subsistence and Environmental Change. Frontiers in Ecology and Evolution 7(313).
  - Guiry EJ, Beglane F, Szpak P, Schulting R, McCormick F, Richards MP. 2018. Anthropogenic changes to the Holocene nitrogen cycle in Ireland. Science Advances 4(6).
  - Guiry EJ, Needs-Howarth S, Friedland KD, Hawkins AL, Szpak P, Macdonald R, Courtemanche M, Holm E, Richards MP. 2016a. Lake Ontario salmon (Salmo salar) were not migratory: a long-standing historical debate solved through stable isotope analysis. Scientific Reports 6.
  - Guiry EJ, Szpak P, Richards MP. 2016b. Effects of lipid extraction and ultrafiltration on stable carbon and nitrogen isotopic compositions of fish bone collagen. Rapid Communications in Mass Spectrometry 30(13):1591-1600.
  - Hadley KR, Douglas MS, Blais JM, Smol JP. 2010. Nutrient enrichment in the High Arctic associated with Thule Inuit whalers: a paleolimnological investigation from Ellesmere Island (Nunavut, Canada). Hydrobiologia 649(1):129-138.
- Hawkins AL, Needs-Howarth S, Orchard TJ, Guiry EJ. 2019. Beyond the local fishing hole: A preliminary study of pan-regional fishing in southern Ontario (ca. 1000 CE to 1750 CE). Journal of Archaeological Science: Reports 24:856-868.
- Head CG. 1975. Introduction to forest exploitation in nineteenth-century Ontario. In: Wood JD, editor. Perspectives on Landscape and Settlement in Nineteenth-Century Ontario:

  McGill-Queen's Press-MQUP. p 78-112.

- Heaton TH. 1986. Isotopic studies of nitrogen pollution in the hydrosphere and atmosphere: a review. Chemical Geology: Isotope Geoscience Section 59:87-102.
- Heckenberger MJ, Kuikuro A, Kuikuro UT, Russell JC, Schmidt M, Fausto C, Franchetto B. 2003. Amazonia 1492: pristine forest or cultural parkland? Science 301(5640):1710-1714.
- Hobson KA, Clark RG. 1992. Assessing Avian Diets Using Stable Isotopes I: Turnover of 13C in Tissues. The Condor 94(1):181-188.

816

824 825

828

829

830

- Hodell DA, Schelske CL. 1998. Production, sedimentation, and isotopic composition of organic matter in Lake Ontario. Limnology and Oceanography 43(2):200-214.
  - Hodell DA, Schelske CL, Fahnenstiel GL, Robbins LL. 1998. Biologically induced calcite and its isotopic composition in Lake Ontario. Limnology and Oceanography 43(2):187-199.
- Hoffman JC, Kelly JR, Peterson GS, Cotter AM, Starry MA, Sierszen ME. 2012. Using δ15N in fish larvae as an indicator of watershed sources of anthropogenic nitrogen: response at multiple spatial scales. Estuaries and coasts 35(6):1453-1467.
- Holm E, Mandrak NE, Burridge ME. 2009. The ROM field guide to freshwater fishes of Ontario.
  Toronto: Royal Ontario Museum.
- Holmes NT. 2006. The importance of long- term data sets in science and river management.

  Aquatic Conservation: Marine and Freshwater Ecosystems 16(4):329-333.
  - Humphries P, Winemiller KO. 2009. Historical impacts on river fauna, shifting baselines, and challenges for restoration. BioScience 59(8):673-684.
- Jeffers ES, Nogué S, Willis KJ. 2015. The role of palaeoecological records in assessing ecosystem services. Quaternary Science Reviews 112:17-32.
  - Jenny J-P, Koirala S, Gregory-Eaves I, Francus P, Niemann C, Ahrens B, Brovkin V, Baud A, Ojala AEK, Normandeau A and others. 2019. Human and climate global-scale imprint on sediment transfer during the Holocene. Proceedings of the National Academy of Sciences 116(46):22972-22976.
- Keizer PS, Gajewski K, McLeman R. 2015. Forest dynamics in relation to multi-decadal late-Holocene climatic variability, eastern Ontario, Canada. Review of Palaeobotany and Palynology 219:106-115.
- Kelly K. 1971. Wheat farming in Simcoe county in the mid-nineteenth century. Canadian Geographer 15(2):95-112.
- Kelly K. 1973. Notes on a type of mixed farming practised in Ontario during the early nineteenth century. Canadian Geographer/Le Géographe canadien 17(3):205-219.
- Kelly K. 1975. The impact of nineteenth-century agricultural settlement on the land. In: Wood JD, editor. Perspectives on Landscape and Settlement in Nineteenth-Century Ontario.
  Ottawa: McClelland and Stewart. p 64-77.
- Kendall C. 1998. Tracing nitrogen sources and cycling in catchments. Isotope tracers in catchment hydrology: Elsevier. p 519-576.
- Kendall C, Elliott EM, Wankel SD. 2007. Tracing anthropogenic inputs of nitrogen to ecosystems. Stable isotopes in ecology and environmental science 2:375-449.
- Kintigh KW, Altschul JH, Beaudry MC, Drennan RD, Kinzig AP, Kohler TA, Limp WF,
  Maschner HD, Michener WK, Pauketat TR. 2014. Grand challenges for archaeology.
  American Antiquity 79(1):5-24.
- Kiriluk RM, Servos MR, Whittle DM, Cabana G, Rasmussen JB. 1995. Using ratios of stable nitrogen and carbon isotopes to characterize the biomagnification of DDE, mirex, and

- PCB in a Lake Ontario pelagic food web. Canadian Journal of Fisheries and Aquatic Sciences 52(12):2660-2674.
- Knowles R. 1982. Denitrification. Microbiological reviews 46(1):43.

862

863

864

865

866

867

868

873

874

875

876

877

878

879

889

890

- Laird KR, Haig HA, Ma S, Kingsbury MV, Brown TA, Lewis C, Oglesby RJ, Cumming BF.
  2012. Expanded spatial extent of the Medieval Climate Anomaly revealed in lakesediment records across the boreal region in northwest Ontario. Global change biology
  18(9):2869-2881.
- Lake JL, McKinney RA, Osterman FA, Pruell RJ, Kiddon J, Ryba SA, Libby AD. 2001. Stable nitrogen isotopes as indicators of anthropogenic activities in small freshwater systems. Canadian Journal of Fisheries and Aquatic Sciences 58(5):870-878.
  - Lean D, Knowles R. 1987. Nitrogen transformations in Lake Ontario. Canadian Journal of Fisheries and Aquatic Sciences 44(12):2133-2143.
  - Leggett M, Johannsson O, Hesslein R, Dixon D, Taylor W, Servos M. 2000. Influence of inorganic nitrogen cycling on the δ15N of Lake Ontario biota. Canadian Journal of Fisheries and Aquatic Sciences 57(7):1489-1496.
  - Lewis FD. 2001. Farm settlement with imperfect capital markets: a life-cycle application to Upper Canada, 1826–1851. Canadian Journal of Economics 34(1):174-195.
  - Lewis SL, Maslin MA. 2015. Defining the anthropocene. Nature 519(7542):171-180.
- Lower ARM, Carrothers WA, Saunders SA. 1938. The North American assault on the Canadian forest: a history of the lumber trade between Canada and the United States: Toronto, The Ryerson Press; New Haven, Yale University Press; [etc., etc.] for the Carnegie Endowment for International Peace, Division of Economics and History.
  - Lu Y, Meyers PA, Johengen TH, Eadie BJ, Robbins JA, Han H. 2010. δ 15 N values in Lake Erie sediments as indicators of nitrogen biogeochemical dynamics during cultural eutrophication. Chemical Geology 273(1):1-7.
  - Lu YH, Meyers PA, Robbins JA, Eadie BJ, Hawley N, Ji KH. 2014. Sensitivity of sediment geochemical proxies to coring location and corer type in a large lake: Implications for paleolimnological reconstruction. Geochemistry, Geophysics, Geosystems 15(5):1960-1976.
- Lumb CE, Johnson TB. 2012. Retrospective growth analysis of lake whitefish (Coregonus clupeaformis) in Lakes Erie and Ontario, 1954-2003. Advances in Limnology:429-454.
- Lyman RL, Cannon KP. 2004. Zooarchaeology and conservation biology: University of Utah Press.
- McAndrews J, Boyko M. Dating recent sediment in Lake Ontario by correlation with a varvedated pollen diagram; 1972. p 1265-1274.
- McClain ME, Boyer EW, Dent CL, Gergel SE, Grimm NB, Groffman PM, Hart SC, Harvey JW, Johnston CA, Mayorga E. 2003. Biogeochemical hot spots and hot moments at the interface of terrestrial and aquatic ecosystems. Ecosystems 6(4):301-312.
  - McFadden MA, Mullins HT, Patterson WP, Anderson WT. 2004. Paleoproductivity of eastern Lake Ontario over the past 10,000 years. Limnology and Oceanography 49(5):1570-1581.
- McFadden MA, Patterson WP, Mullins HT, Anderson WT. 2005. Multi-proxy approach to longand short-term Holocene climate-change: evidence from eastern Lake Ontario. Journal of Paleolimnology 33(3):371-391.
- McIlwraith TF. 1976. Freight capacity and utilization of the Erie and Great Lakes canals before 1850. The Journal of Economic History 36(4):852-877.

- McLauchlan KK, Williams JJ, Engstrom DR. 2013. Nutrient cycling in the palaeorecord: Fluxes from terrestrial to aquatic ecosystems. The Holocene 23(11):1635-1643.
- Meyers PA. 2003. Applications of organic geochemistry to paleolimnological reconstructions: a summary of examples from the Laurentian Great Lakes. Organic geochemistry 34(2):261-289.

906

907

910

911

912 913

914

915

916

917

918

919

920

921

922

923

924

925

- Mills E, Casselman J, Dermott R, Fitzsimons J, Gal G, Holeck K, Hoyle J, Johannsson O, Lantry B, Makarewicz J. 2003. Lake Ontario: food web dynamics in a changing ecosystem (1970-2000). Canadian Journal of Fisheries and Aquatic Sciences 60(4):471-490.
  - Morrissey CA, Boldt A, Mapstone A, Newton J, Ormerod SJ. 2013. Stable isotopes as indicators of wastewater effects on the macroinvertebrates of urban rivers. Hydrobiologia 700(1):231-244.
- Mullins HT, Patterson WP, Teece MA, Burnett AW. 2011. Holocene climate and environmental change in central New York (USA). Journal of Paleolimnology 45(2):243-256.
  - Mumby JA, Larocque SM, Johnson TB, Stewart TJ, Fitzsimons JD, Weidel BC, Walsh MG, Lantry JR, Yuille MJ, Fisk AT. 2018. Diet and trophic niche space and overlap of Lake Ontario salmonid species using stable isotopes and stomach contents. Journal of Great Lakes Research 44(6):1383-1392.
  - Munoz SE, Gajewski K. 2010. Distinguishing prehistoric human influence on late-Holocene forests in southern Ontario, Canada. The Holocene 20(6):967-981.
  - Paquette N, Gajewski K. 2013. Climatic change causes abrupt changes in forest composition, inferred from a high-resolution pollen record, southwestern Quebec, Canada. Quaternary Science Reviews 75:169-180.
  - Pennock JR, Velinsky DJ, Ludlam JM, Sharp JH, Fogel ML. 1996. Isotopic fractionation of ammonium and nitrate during uptake by Skeletonema costatum: Implications for δ15N dynamics under bloom conditions. Limnology and Oceanography 41(3):451-459.
  - Perga M-E. 2010. Potential of  $\delta$  13 C and  $\delta$  15 N of cladoceran subfossil exoskeletons for paleoecological studies. Journal of Paleolimnology 44(2):387-395.
  - Perga ME, Desmet M, Enters D, Reyss JL. 2010. A century of bottom-up-and top-down-driven changes on a lake planktonic food web: A paleoecological and paleoisotopic study of Lake Annecy, France. Limnology and Oceanography 55(2):803–816.
- Post DM. 2002. Using stable isotopes to estimate trophic position: models, methods, and assumptions. Ecology 83(3):703-718.
- Rush SA, Paterson G, Johnson TB, Drouillard KG, Haffner GD, Hebert CE, Arts MT,
  McGoldrick DJ, Backus SM, Lantry BF. 2012. Long- term impacts of invasive species
  on a native top predator in a large lake system. Freshwater Biology 57(11):2342-2355.
- Schelske CL. 1991. Historical nutrient enrichment of Lake Ontario: paleolimnological evidence.
  Canadian Journal of Fisheries and Aquatic Sciences 48(8):1529-1538.
- Schelske CL, Hodell DA. 1991a. Recent changes in productivity and climate of Lake Ontario detected by isotopic analysis of sediments. Limnology and Oceanography 36(5):961-975.
- Schelske CL, Hodell DA. 1991b. Recent changes in productivity and climate of Lake Ontario detected by isotopic analysis of sediments. Limnology and Oceanography 36(3):961-975.
- Schelske CL, Hodell DA. 1995. Using carbon isotopes of bulk sedimentary organic matter to reconstruct the history of nutrient loading and eutrophication in Lake Erie. Limnology and Oceanography 40(5):918-929.

- Schelske CL, Robbins JA, Gardner WS, Conley DJ, Bourbonniere RA. 1988. Sediment record of biogeochemical responses to anthropogenic perturbations of nutrient cycles in Lake Ontario. Canadian Journal of Fisheries and Aquatic Sciences 45(7):1291-1303.
- Schelske CL, Stoermer EF, Conley DJ, Robbins JA, Glover RM. 1983. Early Eutrophication in the Lower Great Lakes: New Evidence from Biogenic Silica in Sediments. Science 222(4621):320-322.
- Schilder J, van Hardenbroek M, Bodelier P, Kirilova EP, Leuenberger M, Lotter AF, Heiri O. 2017. Trophic state changes can affect the importance of methane-derived carbon in aquatic food webs. Proceedings of the Royal Society B: Biological Sciences 284(1857):20170278.
- Schindler DW, Dillon PJ, Schreier H. 2006. A review of anthropogenic sources of nitrogen and their effects on Canadian aquatic ecosystems. Biogeochemistry 79(1-2):25-44.

954

955

956 957

958

959

960

961

962

963

964

965

966

969

970

971

972 973

- Sebilo M, Billen G, Mayer B, Billiou D, Grably M, Garnier J, Mariotti A. 2006. Assessing nitrification and denitrification in the Seine River and estuary using chemical and isotopic techniques. Ecosystems 9(4):564-577.
- Smith VH, Schindler DW. 2009. Eutrophication science: where do we go from here? Trends in Ecology & Evolution 24(4):201-207.
- Stager JC, Cumming BF, Laird KR, Garrigan-Piela A, Pederson N, Wiltse B, Lane CS, Nester J, Ruzmaikin A. 2017. A 1600-year diatom record of hydroclimate variability from Wolf Lake, New York. The Holocene 27(2):246-257.
- Sterner RW, Ostrom P, Ostrom NE, Klump JV, Steinman AD, Dreelin EA, Vander Zanden MJ, Fisk AT. 2017. Grand challenges for research in the Laurentian Great Lakes. Limnology and Oceanography 62(6):2510-2523.
- Stewart TJ, Rudstam L, Watkins J, Johnson TB, Weidel B, Koops MA. 2016. Research needs to better understand Lake Ontario ecosystem function: A workshop summary. Journal of Great Lakes Research 42(1):1-5.
- Szpak P. 2011. Fish bone chemistry and ultrastructure: implications for taphonomy and stable isotope analysis. Journal of Archaeological Science 38(12):3358-3372.
  - Szpak P. 2014. Complexities of nitrogen isotope biogeochemistry in plant-soil systems: implications for the study of ancient agricultural and animal management practices. Frontiers in plant science 5.
    - Szpak P, Buckley M, Darwent CM, Richards MP. 2018. Long- term ecological changes in marine mammals driven by recent warming in northwestern Alaska. Global change biology 24(1):490-503.
- 975 Szpak P, Krippner K, Richards M. 2017a. Effects of Sodium Hydroxide Treatment and 976 Ultrafiltration on the Removal of Humic Contaminants from Archaeological Bone. 977 International Journal of Osteoarchaeology 27(6):1070-1077.
- Szpak P, Metcalfe JZ, Macdonald RA. 2017b. Best practices for calibrating and reporting stable isotope measurements in archaeology. Journal of Archaeological Science: Reports 13:609-616.
- Talbot MR. 2001. Nitrogen Isotopes in Palaeolimnology. In: Last WM, Smol JP, editors.
  Tracking Environmental Change Using Lake Sediments: Physical and Geochemical
  Methods. Dordrecht: Springer Netherlands. p 401-439.
- Trueman CN, Moore A. 2007. Use of the stable isotope composition of fish scales for monitoring aquatic ecosystems. Terrestrial Ecology 1:145-161.

- van der Merwe NJ, Williamson RF, Pfeiffer S, Thomas SC, Allegretto KO. 2003. The Moatfield ossuary: isotopic dietary analysis of an Iroquoian community, using dental tissue. Journal of Anthropological Archaeology 22(3):245-261.
- van Hardenbroek M, Heiri O, Grey J, Bodelier PL, Verbruggen F, Lotter AF. 2010. Fossil chironomid δ 13 C as a proxy for past methanogenic contribution to benthic food webs in lakes? Journal of Paleolimnology 43(2):235-245.
- Warrick G. 2008. A population history of the Huron-Petun, AD 500-1650. Cambridge, UK: Cambridge University Press.
- Williamson RF. 2014. The Archaeological History of the Wendat to AD 1651: An Overview.
  Ontario Archaeology 94:3-64.
- Wood D. 2000. Making Ontario: McGill-Queen's Press-MQUP.

- Yang J-R, Duthie HC, Delorme LD. 1993. Reconstruction of the recent environmental history of
   Hamilton Harbour (Lake Ontario, Canada) from analysis of siliceous microfossils.
   Journal of Great Lakes Research 19(1):55-71.
- Yuille MJ, Fisk AT, Stewart T, Johnson TB. 2015. Evaluation of Lake Ontario salmonid niche space overlap using stable isotopes. Journal of Great Lakes Research 41(3):934-940.