



Investigating Maternal Brain Alterations in Preeclampsia

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**Investigating maternal brain alterations in preeclampsia: the need for a
multidisciplinary effort**

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Abbreviation list

40

Blood brain barrier (BBB); Blood pressure (BP); Cerebral blood flow (CBF); Cerebral
42 vascular resistance (CVR); Cerebrospinal fluid (CSF); Gamma amino butyric acid
(GABA); Human pluripotent stem cells (hPSCs); Junctional Adhesion Molecules (JAMs);
44 Lipopolysaccharide (LPS); Magnesium sulphate (MgSO₄); Magnetic resonance imaging
(MRI); Magnetic resonance spectroscopy (MRS); Magnetic resonance spectroscopy
46 focused on hydrogen metabolites (H-MRS); Magnetic resonance spectroscopy focused on
phosphorus metabolites (P-MRS); Neurofilament light chain (NfL); Neuron Specific
48 Enolase (NSE); Pentylentetrazole (PTZ); Posterior reversible encephalopathy syndrome
(PRES); Reduced uteroplacental perfusion pressure (RUPP); Reduced uteroplacental
50 perfusion pressure plus high cholesterol diet (RUPP+HC); S100 calcium-binding protein B
(S100B); Transendothelial electrical resistance (TEER); White matter lesions (WML)

52 **Abstract:**

54 **Porpuose of Review:** To provide insight into the mechanisms underlying cerebral pathophysiology in preeclampsia and to highlight possible methods for evaluation, screening and surveillance of cerebral complications in the condition.

56 **Recent Findings:** Every 12 minutes, a woman dies as a consequence of preeclampsia with cerebral complications, such as eclampsia, among the most common causes. The incidence
58 of eclampsia has been estimated to vary from one in 100 to one in 2000 deliveries, with the highest incidences in low-income countries. The pathophysiology of eclampsia remains
60 enigmatic. The increased blood pressure cannot be the only underlying cause, since some cases of eclampsia arise without simultaneous hypertension. Evaluation of brain alterations
62 in preeclampsia and eclampsia is challenging and demands a multidisciplinary collaboration, since no single method can accurately and fully describe how preeclampsia
64 affects the brain.

Summary: Cerebral complications of preeclampsia are a significant factor in maternal morbidity and mortality worldwide. No single method can accurately describe the full
66 picture of how preeclampsia affects the brain vasculature and parenchyma. We recommend a multinational effort to overcome, not only the issue of limited sample availability, but
68 also, to optimize the quality of research.

70

72 **Keywords:** preeclampsia; eclampsia; brain complications; brain blood barrier; preclinical studies; biomarkers; brain imaging

Introduction

Preeclampsia has classically been characterized by new onset of hypertension and proteinuria after 20 weeks of gestation [1]. However, in several new guidelines, proteinuria is not mandatory for diagnosis if the onset of hypertension is accompanied by other signs of organ impairment including thrombocytopenia, renal insufficiency, impaired liver function, pulmonary edema, fetal growth restriction or seizures (eclampsia) [2, 3]. The majority of maternal deaths related to preeclampsia can be avoided by providing timely and effective care and delivery to high-risk women [4]. Thus, optimization of health care for women during pregnancy to prevent and treat preeclampsia, both in high and low-income countries, is a necessary step towards achievement of the Millennium Development Goals [5].

Eclampsia is defined as tonic-clonic seizures in the presence of preeclampsia and in the absence of other underlying causes of seizures [6]. In cases of recurrent seizures, there is a higher risk of severe hypoxia, aspiration pneumonia, maternal injury and status epilepticus. [7]. Although eclampsia has been defined as a severe complication of preeclampsia, it can occur even in the absence of hypertension [8]. Eclampsia occurs in about 1 in 2000 deliveries in high income countries, while in low and middle income countries it is estimated that the incidence of eclampsia varies from 1 in 100 to 1 in 1700 deliveries [9] (Figure 1).

Currently, there is a gap in knowledge regarding the underlying pathophysiology of preeclampsia and eclampsia. This gap in knowledge impairs the development of tools for the diagnosis and management of women with neurological impairment such as eclampsia or intracerebral edema. Research within the field has been hampered by; i) the low incidence of eclampsia in high income countries where most research is conducted; ii) the fact that preeclampsia is a uniquely human condition; iii) poor access to imaging techniques for evaluation of maternal brain function and iii) the limited applicability of *in vitro* models.

This review aims to present the current knowledge of preeclampsia and its neurological effects and to highlight the need for a multidisciplinary approach, including

104 utility of preclinical models, clinical tools for evaluation and analysing the impact of
findings from clinical studies, to better understand the pathophysiology of the condition.

106

Diagnosis and treatment of eclampsia

108

In pregnant women, there are no imaging criteria or circulating biomarkers to aid in
110 the prediction of eclampsia, and even clinical predictors may have poor predictive ability
for eclampsia. The diagnosis can be challenging if the woman is not diagnosed with
112 preeclampsia before onset of seizures or if there are other underlying disorders that can
possibly contribute to seizures. In addition, seizures are often unexpected, even in the cases
114 where women present with preeclampsia before onset of eclampsia.

The cornerstones in the treatment of eclampsia are prevention of maternal injury,
116 support of respiratory and cardiovascular function, prevention of recurrent convulsions and
reduction of blood pressure (BP) to a safe range [10]. Systolic BP control is essential in
118 avoiding hemorrhagic stroke and should be kept below 160 mmHg [11, 12]. Treatment with
magnesium sulphate (MgSO₄) is indicated as clinical management for manifest eclampsia
120 to avoid recurrent seizures [13]. In women with eclampsia, MgSO₄ treatment reduces the
risk of recurrent fits and maternal death by 59% and 38%, respectively [10]. Furthermore,
122 treatment with MgSO₄ has a protective effect against developing eclampsia, in particular in
women with severe preeclampsia or with imminent signs of eclampsia (blurred vision,
124 severe epigastric pain or headache) [13]. Despite being the drug of choice for preventing
eclampsia and maternal death, the mechanism of action of MgSO₄ is not well understood.
126 Suggested mechanisms include reducing blood brain barrier (BBB) permeability, reducing
neuroinflammation, or modulation of gamma amino butyric acid (GABA) receptor
128 activation [14].

130 Cerebral alterations after preeclampsia and eclampsia

132 Cerebral imaging of women with eclampsia often shows a condition defined as
posterior reversible encephalopathy syndrome (PRES), a type of cerebral edema thought to
134 be reversible without longterm effects. However, lately, there have been questions about

136 the actual reversibility of the cerebral effects of eclampsia, or severe preeclampsia, since
138 women who have experienced preeclampsia previously are at higher risk of stroke, vascular
140 dementia, epilepsy and cognitive failure months to years after pregnancy [15-17]. Magnetic
142 resonance imaging (MRI) studies have revealed changes in both morphology and function
144 of the brain in women with preeclampsia and eclampsia. For example, women with a
146 previous pregnancy complicated by eclampsia or severe preeclampsia have an increased
148 number of white matter lesions (WML) several years after the event [18, 19]. These lesions
are areas of hyperintensity on T2 weighted MRI brain scans that are known to correlate
with cognitive decline and dementia [20, 21]. The distribution of WML does not appear to
correspond to areas most often affected in PRES, a finding which argues against the notion
of a direct causal pathway [22]. A recent study did, however, show reduced gray matter
volume in posterior localizations in the brain in women with a history of preeclampsia and
current hypertension [23]. In addition, PRES is not always restricted to the posterior
regions of the brain but can also be located in the temporal or frontal regions [24].

Others also report reduced cortical volume or total brain volume in women with a
150 history of preeclampsia [25, 26]. In the study by Mielke et al [27], women with a history of
pregnancy hypertensive disease performed worse in a cognitive test measuring processing
152 speed and had greater brain atrophy compared to women with previous normal pregnancies.
These recordings were made decades after the affected pregnancy, and differences
154 remained statistically significant even after adjustment for traditional cardiovascular risk
factors [25]. More recently, a meta-analysis of cognitive function after preeclampsia, found
156 a correlation between preeclampsia and later subjective, but not objective, impairment of
cognition. However the authors also stated that high quality studies are lacking in this field
158 and that most studies did not correct for confounders [28].

160 **Cerebrovascular blood flow in preeclampsia and eclampsia**

162 The etiology of eclampsia and cerebral complications in relation to preeclampsia is
not well understood, but it is thought to be the result of impaired regulation of
164 cerebrovascular function resulting in mainly vasogenic edema, but also partly cytotoxic
edema, predominately in the parieto-occipital regions of the brain. The underlying

166 mechanism might be cerebral vasoconstriction, impaired autoregulation with forced
dilatation of cerebral arteries and/or endothelial dysfunction [8, 29, 30].

168 Studies employing animal models of preeclampsia have demonstrated that cerebral
vasogenic edema is the result of impaired autoregulation of cerebral blood flow and
170 increased BBB permeability [31]. However, since preeclampsia is a condition unique to
humans, findings from animal studies with induced preeclampsia can be difficult to
172 interpret and translate to humans.

The original basic concept of the underlying cause of PRES, cerebral vasoconstriction,
174 is based on findings where angiography had shown caliber-changes in the cerebral arteries
where vasoconstriction was thought to occur in response to hypertension with
176 autoregulatory compensation, resulting in hypoxia, endothelial dysfunction and subsequent
vasogenic and cytotoxic edema [32, 33]. This theory has become less popular in favour for
178 failure of cerebral autoregulation and hyperperfusion, but not abandoned.

Cerebral vascular autoregulation is a physiological mechanism that maintains a
180 relatively constant cerebral blood flow despite changes in cerebral perfusion pressure. The
theory proposed to explain the loss of cerebral vascular autoregulation is based on dilation
182 of brain vessels (decrease in cerebral vascular resistance, CVR) instead of vessel
constriction, in response to increasing systemic blood pressure. This causes increased
184 cerebral blood flow (CBF), increased pressure on the vessel wall which would in turn
causes intracerebral vasogenic edema [34]. However, this process can only occur when the
186 upper limit of the cerebral autoregulation is reached, which is not the case in many women
with eclampsia and preeclampsia with PRES [35, 36]. Consequently, the reason why
188 pregnant women, with blood pressures within the normal range of cerebral autoregulation,
develop neurological symptoms and/or eclampsia remains unexplained [37]. Some studies
190 have shown that it might be the dynamic cerebral autoregulation (i.e continuous rapid
changes in vessel diameter in response to small changes in systemic blood pressure) that is
192 impaired in preeclampsia. This might be one of the explanations as to why eclampsia can
occur at blood pressure levels that are lower than the upper range of cerebral autoregulation
194 [38].

Thus, at present, our knowledge of the pathophysiology of edema and seizures in
196 preeclampsia is incomplete, and further research is required.

198 **Endothelial dysfunction in preeclampsia**

200 Endothelial dysfunction is a systemic pathological state characterized by the loss of
the physiological response of the endothelium to mediators of vasodilation and
202 vasoconstriction derived from both endothelial and non-endothelial origin [39]. This
condition has been reported in women with preeclampsia [40] and is thought to be a
204 generalized condition that targets multiple organs, including the brain. In this review we
focus on the importance of brain endothelial cells, a key component of the BBB, in relation
206 to preeclampsia pathophysiology.

The BBB is a complex structure primarily composed of capillary endothelial cells in
208 conjunction with astrocytes, pericytes and basement membrane. In the BBB, adjacent
endothelial cells are tightly associated via intercellular tight junction complexes that restrict
210 paracellular transport, thereby regulating the cerebral environment [41]. Consequently,
disruption of tight junction function results in loss of BBB integrity and subsequent
212 increased permeability.

The key components of intercellular tight junctions are the transmembrane proteins
214 occludin, claudin and Junctional Adhesion Molecules (JAMs), which form complex strands
that govern the permeability characteristics of the paracellular route [42, 43]. In order to
216 maintain this restricted diffusion pathway, tight junctions are linked to the cytoplasmic
zonula occludens proteins that provide a structural bridge to the actin cytoskeleton.
218 Furthermore, the phenotype of BBB endothelial cells differs from that of peripheral
endothelial cells, since they express higher levels of tight junction proteins, membrane
220 transporters belonging to the ATP-binding cassette and Solute Carrier families, and
metabolic enzymes [44-46]. It is this phenotype that is fundamentally responsible for
222 defining the highly restrictive vascular permeability characteristics of the BBB.

Since preeclampsia with cerebral complications may occur in pregnant women with
224 mild hypertension, or even without diagnosis of hypertension [47], it is accepted that
circulating factors, probably released from the placenta, may target brain endothelial cells
226 and increase BBB permeability, potentially facilitating the onset of edema and seizures
(Figure 2). In support of this hypothesis, Warrington et al [48] reported that pregnant rats

228 with induced placental ischemia demonstrated increased permeability of the cerebral
vasculature. Thus, factors released from the ischemic placenta in preeclampsia might
230 sensitize the cerebral vasculature to changes in blood pressure, which might enhance BBB
permeability.

232 Both Cipolla et al [31] and Johnson et al 2014 [14] have reported a hypertensive
pregnancy-dependent significant increase in BBB permeability to sodium fluorescein (376
234 Da) in a rat model of severe preeclampsia, whilst Cipolla and Kraig [31], but not Johnson et
al [14], also showed increased permeability of Texas red dextran (70 kDa). The increased
236 BBB permeability in this model may be associated with tight junction disruption, but no
direct evidence has yet been reported. Furthermore, rat studies investigating the effect of
238 preeclampsia on brain physiology postpartum report that decreased expression of the tight
junction protein occludin in the posterior cortex was associated with edema [49].

240 Several reports have described a change in circulating concentrations of cytokines in
women who developed preeclampsia, i.e. higher proinflammatory and lower anti-
242 inflammatory concentrations compared to normal pregnancies [50, 51]. However, little is
known about circulating concentrations of these cytokines in women who experienced
244 brain complications such as eclampsia, although circulating TNF- α may contribute to
cerebral edema by increasing BBB permeability [52]. Furthermore, in *in vitro* studies the
246 increase in BBB permeability by plasma from women with preeclampsia could be
prevented by inhibition of vascular endothelial growth factor signaling [53]. Future studies
248 should focus on the impact of the increased inflammatory state on the brain vasculature in
preeclampsia.

250

Evaluation of brain function in women with preeclampsia or eclampsia

252

Preclinical models

254 *Animal models*

256 Cerebral complications in preeclampsia are difficult to simulate in *in vivo* models.
Classical models for preeclampsia, such as the reduced uteroplacental perfusion pressure
258 (RUPP) model have been combined with high cholesterol diet (RUPP+HC) in order to

mimic severe preeclampsia [14]. This RUPP+HC model results in high BP and reduced placental and pup weights. To evaluate eclampsia in this RUPP+HC model, seizures are induced by administration of the neuroexcitatory agent pentylenetetrazole (PTZ). In this study, seizure threshold to PTZ was decreased, indicating that preeclampsia had a harmful effect on the brain. Furthermore, in the same RUPP+HC model, a lower percentage water content in the posterior cerebral cortex, high *in vivo* BBB permeability to sodium fluorescein and high microglia activation were demonstrated RUPP+HC rats compared to late pregnant controls [14]. The latter study also showed administration of MgSO₄ reversed the above effects.

More recently, another rat model of eclampsia, based on administration of lipopolysaccharide (LPS) plus PTZ to pregnant rats, has been validated by different groups [54, 14, 55]. This model demonstrates not only preeclampsia-like syndrome plus seizures, but neuroinflammation, brain edema and high levels of proinflammatory cytokines in peripheral blood and in cerebrospinal fluid. These abnormalities can be prevented by MgSO₄. Interestingly, placental ischemia hastened the onset of seizures compared to pregnant controls but had no effect on seizure duration. Other available rat models of hypertension in pregnancy have not been used in studies of maternal brain alteration.

Despite validation of available eclampsia models, rat models have limited applicability in terms of defining whether the brain regions affected in rodents correspond to the affected areas in human. In addition, since preeclampsia is a uniquely human condition, an animal model can never truly reflect the effects of preeclampsia on the human brain. Consequently, there is a need for more studies to better understand how cerebral complications arise during severe preeclampsia [31].

In vitro models

Since the preeclampsia-associated modulation of BBB permeability is a multifactorial phenomenon involving alterations in the restrictive characteristics of tight junction complexes and intracellular signaling events, *in vitro* models of the BBB can prove useful in characterizing these alterations. Ideally, the models should be functionally reproducible, retaining key characteristics of the *in vivo* BBB, including high

290 transendothelial electrical resistance (TEER), low permeability, and expression of
functional receptors and associated signaling and endocytotic pathways, ATP-binding
292 cassette, Solute Carrier transporters, and tight junction proteins. A summary of the
available models is presented in Table 1.

294 There is very rarely healthy human brain tissue available to generate BBB due to
obvious ethical issues. However primary human brain endothelial cells are commercially
296 available. Immortalised cell lines are an alternative to primary cell cultures, although
immortalization has a negative impact on normal cell physiology. Of all the human-based
298 models [56, 57] the hCMEC/d3 cell line has consistently proved to be the most reliable in
terms of phenotype and relevance to studying BBB function [58-60], whilst in recent years,
300 the use of human pluripotent stem cells (hPSCs) for developing *in vitro* models of the
human BBB appears to be extremely promising [61].

302 Animal-derived BBB models have obvious advantages over human-based models in
terms of tissue availability and versatility, and brain tissue from rodents has been widely
304 employed. More robust BBB models, based on bovine and porcine primary endothelial
cells, demonstrate high TEER [62-66] low permeability of small tracer molecules [64, 62,
306 63, 66], expression of phenotypical proteins [62, 67, 68, 65] and responsiveness to
endogenous mediators and xenobiotics [69, 70]. *In vitro* BBB models have also been
308 generated using immortalized rodent, bovine and porcine endothelial cell lines, which can
express tight junction proteins and functionally active BBB transporters [71-74, 57, 75].

310 All *in vitro* model systems have limitations (Table 1), and the major considerations
when employing *in vitro* BBB models are optimization of culture conditions and the use of
312 more complex co-culture systems to maintain, or enhance, the *in vivo* BBB phenotype.
These considerations will help create a reliable and reproducible model system in which to
314 study BBB function.

316 **Clinical studies**

Brain Imaging in Preeclampsia

318 Brain imaging is the gold standard for analysis of brain alterations during severe
preeclampsia. These are summarized in Table 2.

320

Transcranial Doppler.

322

Transcranial Doppler ultrasound is a non-invasive technique that can be utilized to
324 measure cerebral blood flow. The middle cerebral artery is the most commonly studied
vessel due to the ease of access via the temporal window. Transcranial Doppler has been
326 used for assessment of cerebrovascular function in clinical settings, including vasospasm in
subarachnoid hemorrhage and stroke. In Transcranial Doppler studies of the middle
328 cerebral artery, cerebral perfusion pressure has been shown to be increased in women with
preeclampsia compared to normal pregnant women [76, 77, 38]. Consequently, Belfort et al
330 proposed that elevated cerebral perfusion pressure may contribute to the pathophysiology
of vasogenic edema and hypertensive encephalopathy in preeclampsia/eclampsia [78].

332 Dynamic cerebral autoregulation, the ability of vessels to respond to subtle changes in
systemic blood pressure by contracting or dilating has been studied in normal pregnancies
334 and in pregnancies complicated by hypertensive disorders, using a combination of
Transcranial Doppler, continuous non-invasive blood pressure monitoring, and continuous
336 end-tidal CO₂ monitoring [79]. Women with preeclampsia have been shown to have
impaired dynamic autoregulation compared to women with a normal pregnancy and,
338 interestingly, this impairment does not correlate to higher systemic blood pressure [38]. A
possible mechanism is that elevated cerebral perfusion pressure and/or impaired dynamic
340 autoregulation may lead to disruption of the endothelium and microstructure of the cerebral
vasculature (barotrauma) and result in cerebral edema and hemorrhage.

342 Compared to the gold standard of angiography, Transcranial Doppler offers the
advantage of decreased expense and lack of radiation exposure. Limitations of Transcranial
344 Doppler include the dependence on the operator for the handheld technique and anatomic
variations including inadequate acoustic windows. Most studies evaluating cerebral blood
346 flow in preeclampsia/eclampsia have focused on sonation of the middle cerebral artery.
However, since most cerebral pathology in preeclampsia/eclampsia involves the posterior
348 cerebral circulation (visual disturbances, cortical blindness, and posterior reversible
encephalopathy) future studies should evaluate the posterior cerebral circulation. Studies
350 monitoring cerebral perfusion pressure and dynamic cerebral autoregulation in both

eclampsia and preeclampsia are lacking, and more efforts are required to determine the
352 influence of these parameters on cerebral complications.

354 *Magnetic Resonance Imaging (MRI)*

356 Magnetic resonance imaging (MRI) produces very high contrast images of soft
tissue, making it the method of choice for imaging the brain and spinal cord. In the context
358 of preeclampsia, MRI can prove useful for detecting increases in water content, e.g. due to
gliosis or vasogenic edema and has been used for estimation of white matter lesion burden,
360 for detection of the PRES, and for cortical volume assessments (Figure 3).

Besides morphology, MRI can be used for investigations of physiology and
362 metabolism. With a diffusion weighted sequence (diffusion weighted imaging), it is
possible to differentiate vasogenic and cytotoxic edema. Blood flow and flow velocity can
364 be measured with phase contrast MR technique [80]. Although various contrast agent-based
techniques are used for assessment of tissue perfusion, they cannot be used in pregnancy.
366 There are however newer MR perfusion techniques that do not employ contrast agents,
namely arterial spin labeling [81] and intravoxel incoherent motion [82], and intravoxel
368 incoherent motion studies report reduced perfusion in a part of the basal ganglia in women
with preeclampsia [83].

370 Tissue metabolism can be investigated with MR spectroscopy, most often focusing
on hydrogen metabolites (H-MRS)[84], but also on phosphorus metabolites (P-MRS) [85]
372 and a P-MRS study reports a reduction in cerebral magnesium levels of the brain in women
with preeclampsia [86]. Using H-MRS, changes in osmolytes were detected in the brains of
374 women with preeclampsia [87, 88], suggesting that brain and plasma osmolality may play a
role in the cerebral edema associated with preeclampsia and eclampsia. To date, the MR
376 spectroscopy technique has rarely been used in women with eclampsia.[89]

Disadvantages with MRI include that it is not as available as ultrasound or CT,
378 examination times are longer, cost is higher, thereby making it difficult to examine
critically ill patients.

380

Cerebral biomarkers

382

In preeclampsia, four cerebral biomarkers have been investigated in the setting of
384 brain alterations: 1) S100B (10.7 kDa monomer; 21 kDa homodimer) from astroglial cells,
2) Neuron Specific Enolase (NSE) (47 kDa) from neurons 3) Neurofilament light chain
386 (NfL) (68 kDa) and 4) tau (six isoforms, 36.8 - 45.9 kDa) from axons. Amongst these,
S100B has been most studied. If these centrally-derived biomarkers, when detected
388 peripherally, are proven to have a high accuracy in diagnosing cerebral complications at
onset or before onset, they could be used to predict or diagnose cerebral complications in
390 preeclampsia.

However, in terms of applicability in the clinical setting, there is still a large gap in
392 knowledge about the utility of circulating cerebral biomarkers in the prediction and
diagnosis of cerebral complications in preeclampsia. Most studies have evaluated women
394 with preeclampsia without cerebral complications. Prospective studies investigating women
with cerebral complications in preeclampsia, such as cerebral edema and/or eclampsia, are
396 needed to verify the accuracy of these biomarkers.

The possible advantage of analyzing circulating biomarkers is that women with
398 cerebral complications, or manifest cerebral signs, could be evaluated and treated according
to their risk of complications. This would facilitate allocation of treatment such as
400 magnesium sulphate, decision about delivery and allocation to the right level of care.
Briefly, we will analyze current available biomarkers in the next sections.

402

S100B in preeclampsia

404

Since S100B is produced by astroglial cells, and in particular in the end-feet
406 surrounding the neurovascular unit, and it is thought that S100B might enter the blood
stream after an isolated BBB injury, even without injury to the brain parenchyma [90]. This
408 is supported by studies examining the loss of BBB integrity in pharmacological studies of
drugs directed to the brain [91].

410 Increased plasma levels of S100B have been reported in women with eclampsia [92],
whilst women with severe preeclampsia had higher plasma levels of S100B compared to
412 women with mild disease [93]. Also, increased plasma concentrations of S100B were

414 correlated with visual disturbances among women with preeclampsia [94] or with cerebral
symptoms [95]. Therefore, S100B seems to be a promising blood-based biomarker for
cerebral impairment in preeclampsia.

416

Neuron Specific Enolase in preeclampsia

418

420 Neuron Specific Enolase (NSE) is found in neurons, red blood cells and in the
neuroendocrine system. NSE in combination with clinical parameters has been
recommended as neurologic prognostication in patients with cardiac arrest and hypoxic
422 ischemic encephalopathy [96].

424 Little information is available on these markers in preeclampsia. Two studies report
that plasma concentrations of NSE were increased in late pregnancy in women developing
preeclampsia [97] and that plasma concentrations of NSE were still increased one year after
426 delivery in women who had preeclampsia compared to healthy controls [98].

NfL and tau in preeclampsia

430 Neurofilament light chain (NfL) and tau are axonal proteins used as biomarkers for
neurodegenerative disease. NfL is released into the cerebrospinal fluid (CSF) and
432 subsequently peripheral blood and might be useful to rule out intracranial pathology in
patients with traumatic brain injury [99]. Serum concentrations of tau have been proven to
434 predict 6 months cerebral outcome after cardiac arrest in a pilot study [100]. Two studies
have shown that concentrations of NfL are increased at the end of pregnancy but before
436 onset of disease in women developing preeclampsia [101, 102] and one of these studies
also showed that tau was increased before onset of disease [102]

438

Concluding remarks

440

442 Evaluation of brain alterations in preeclampsia and eclampsia is challenging and
demands collaboration between experts in the laboratory, imaging specialties and in the
clinical setting. No single method can accurately describe the full picture of how

444 preeclampsia affects the brain vasculature and parenchyma since there are sources of error
no matter which method is employed.

446 Also, different mechanisms of pathophysiology have been reported, and evidence
has been hard to pool since studies usually report findings from only one modality. If a
448 combination of peripheral biomarkers, cerebral imaging and *in vitro* studies support the
same findings, this will strengthen the validity of the results (Figure 4). In future research
450 in preeclampsia and eclampsia, the above methods of evaluation of cerebrovascular
alterations should be combined and outcomes should be focused on both short- and long-
452 term cerebral complications such as eclampsia, PRES, impaired cognitive function and
stroke. If women could be objectively identified as low or high risk of cerebral
454 complications, treatment and support could be directed to the women at highest risk. In
terms of prevention, identification of a biomarker that accurately reflects the risks of
456 eclampsia and other preeclampsia-associated severe cerebral complications, as well as brain
alterations later in life after a pregnancy complicated by preeclampsia, is highly desirable.
458 However, eclampsia is a severe, but rare condition. We recommend a multinational effort to
overcome the issue of limited sample availability, and a translational research approach to
460 optimize the quality of research. Cerebral complications of preeclampsia are a significant
factor in maternal morbidity and mortality worldwide. To reach the 5th WHO Millennium
462 goal for decreased maternal mortality, there is undoubtedly a need for a multidisciplinary
effort to gain increased knowledge of brain vascular alterations in preeclampsia.

464

466

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470

Author's roles

472 CE and LB: Designed and wrote the manuscript. All co-authors included their respective
sections according to expertise. JMR, AKW and JP contributed to the writing of the
474 manuscript and provided a critical revision of its contents. All coauthors approval the final
version of this manuscript.

476 **Figure Legends**

478 **Figure 1: Incidence of eclampsia worldwide.** Figure indicates calculated incidence from
literature described in supplementary Table S1. Calculated incidence per 100.000
480 deliveries.

482 **Figure 2: Current concept of the pathophysiology of brain alterations in preeclampsia
and eclampsia.** **A)** We propose a communication between placental and maternal brain. **B)**
484 The ischemic placenta releases harmful substances, including exosomes, into the maternal
circulation, which in turn reach brain endothelial cells generating damage in the blood brain
486 barrier. **C)** Increased brain endothelial permeability will impair the function of brain
parenchyma.

488

Figure 3. Magnetic resonance images of woman with eclampsia. FLAIR sequence **A)**
490 shows high signal intensity changes compatible with posterior encephalopathy syndrome
(PRES) in bilateral parietooccipital areas as well as in bilateral basal ganglia. Intravoxel
492 incoherent motion (IVIM) images show reduced fast diffusion (D^*), indicating reduced
blood flow velocity **B)** and reduced perfusion fraction (f) indicating reduced blood volume
494 **C).** Changes are more obvious in D^* image. The findings support hypoperfusion in areas
corresponding to the PRES findings.

496

**Figure 4: Approach to the study of brain alterations in preeclampsia and eclampsia
from bench to bedside.** There is a need to develop multidisciplinary approaches to better
understand the pathophysiology of brain alterations associated with preeclampsia and
500 eclampsia. Among others, we have summarized the necessity for further improvements in:
clinical management, epidemiological studies, brain imaging tools, development of
502 prevention strategies such as use of biomarkers; and further preclinical research, including
in vitro and animal model studies. This multidisciplinary approach will result in an iterative
504 process leading to advancement of knowledge in the preeclampsia cerebral vascular field
and will help reduce fetomaternal complications due to brain alterations.

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Table Legends

512 **Table 1. Features of *in vitro* blood-brain barrier models for studies in preeclampsia***

*References within the manuscript

514

516 **Table 2: Features of clinical approaches for studying cerebral complications in preeclampsia.**

*References within the manuscript

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520 **References**

- 522 1. Brown MA, Lindheimer MD, de Swiet M, Assche AV, Moutquin J-M. The
524 Classification and Diagnosis of the Hypertensive Disorders of Pregnancy: Statement
526 from the International Society for the Study of Hypertension in Pregnancy (ISSHP).
Hypertension in Pregnancy. 2001;20:ix-xiv. doi:10.3109/10641950109152635.
- 528 2. ACOG TFOHiP. Hypertension in Pregnancy. Washington: 2013.
- 530 3. Brown MA, Magee LA, Kenny LC, Karumanchi SA, McCarthy FP, Saito S et al. The
528 hypertensive disorders of pregnancy: ISSHP classification, diagnosis &
530 management recommendations for international practice. *Pregnancy Hypertens.*
2018. doi:10.1016/j.preghy.2018.05.004.
- 532 4. Campbell OM, Graham WJ. Strategies for reducing maternal mortality: getting on with
532 what works. *Lancet.* 2006;368:1284-99. doi:10.1016/S0140-6736(06)69381-1.
- 534 5. Say L, Chou D, Gemmill A, Tunçalp O, Moller AB, Daniels J et al. Global causes of
534 maternal death: a WHO systematic analysis. *Lancet Glob Health.* 2014;2(6):e323-
33. doi:10.1016/S2214-109X(14)70227-X.
- 536 6. Mol BWJ, Roberts CT, Thangaratinam S, Magee LA, de Groot CJM, Hofmeyr GJ. Pre-
536 eclampsia. *The Lancet.* 2016;387:999-1011. doi:10.1016/S0140-6736(15)00070-7.
- 538 7. Sibai BM. Magnesium sulfate prophylaxis in preeclampsia: Lessons learned from recent
538 trials. *American Journal of Obstetrics and Gynecology.* 2004;190:1520-6.
540 doi:10.1016/j.ajog.2003.12.057.
- 542 8. Sibai B, Dekker G, Kupferminc M. Pre-eclampsia. *Lancet.* 2005;365(9461):785-99.
542 doi:10.1016/S0140-6736(05)17987-2.
- 544 9. Duley L. The global impact of pre-eclampsia and eclampsia. *Seminars in perinatology.*
544 2009;33(3):130-7. doi:10.1053/j.semperi.2009.02.010.
- 546 10. Sibai BM. Diagnosis, prevention, and management of eclampsia. *Obstet Gynecol.*
546 2005;105(2):402-10.
- 548 11. Basit S, Wohlfahrt J, Boyd HA. Pre-eclampsia and risk of dementia later in life:
548 nationwide cohort study. *Bmj.* 2018;k4109. doi:10.1136/bmj.k4109.
- 550 12. Brussé I, Duvekot J, Jongerling J, Steegers E, De Koning I. Impaired maternal
550 cognitive functioning after pregnancies complicated by severe pre-eclampsia: A
552 pilot case-control study. *Acta Obstetrica et Gynecologica Scandinavica.*
2008;87:408-12. doi:10.1080/00016340801915127.
- 554 13. Duley L. Do women with pre-eclampsia, and their babies, benefit from magnesium
554 sulphate? The Magpie Trial: A randomised placebo-controlled trial. *Lancet.*
2002;359:1877-90. doi:10.1016/S0140-6736(02)08778-0.
- 556 14. Johnson AC, Tremble SM, Chan SL, Moseley J, LaMarca B, Nagle KJ et al.
556 Magnesium sulfate treatment reverses seizure susceptibility and decreases
558 neuroinflammation in a rat model of severe preeclampsia. *PLoS One.*
2014;9(11):e113670. doi:10.1371/journal.pone.0113670.
- 560 15. Andolf EG, Sydsjo GC, Bladh MK, Berg G, Sharma S. Hypertensive disorders in
560 pregnancy and later dementia: a Swedish National Register Study. *Acta obstetrica
562 et gynecologica Scandinavica.* 2017;96(4):464-71. doi:10.1111/aogs.13096.
- 564 16. Basit S, Wohlfahrt J, Boyd HA. Pre-eclampsia and risk of dementia later in life:
564 nationwide cohort study. *BMJ.* 2018;363:k4109. doi:10.1136/bmj.k4109.

- 566 17. Nerenberg KA, Park AL, Vigod SN, Saposnik G, Berger H, Hladunewich MA et al.
Long-term Risk of a Seizure Disorder After Eclampsia. *Obstet Gynecol.* 2017;130(6):1327-33. doi:10.1097/AOG.0000000000002364.
- 568 18. Aukes AM, De Groot JC, Wiegman MJ, Aarnoudse JG, Sanwkarja GS, Zeeman GG.
Long-term cerebral imaging after pre-eclampsia. *BJOG.* 2012;119(9):1117-22.
570 doi:10.1111/j.1471-0528.2012.03406.x.
- 572 19. Aukes AM, de Groot JC, Aarnoudse JG, Zeeman GG. Brain lesions several years after
eclampsia. *Am J Obstet Gynecol.* 2009;200(5):504 e1-5.
doi:10.1016/j.ajog.2008.12.033
- 574 S0002-9378(08)02439-3 [pii].
- 576 20. Enzinger C, Fazekas F, Ropele S, Schmidt R. Progression of cerebral white matter
lesions - Clinical and radiological considerations. *Journal of the Neurological
Sciences.* 2007;257:5-10. doi:10.1016/j.jns.2007.01.018.
- 578 21. Prins ND vDE, den Heijer T, et al. Cerebral White Matter Lesions and the Risk of
Dementia. *Arch Neurol.* 2004;61:1531-4.
- 580 22. Wiegman MJ, Zeeman GG, Aukes AM, Bolte AC, Faas MM, Aarnoudse JG et al.
Regional distribution of cerebral white matter lesions years after preeclampsia and
582 eclampsia. *Obstet Gynecol.* 2014;123(4):790-5.
doi:10.1097/AOG.000000000000162
- 584 00006250-201404000-00010 [pii].
- 586 23. Raman MR, Tosakulwong N, Zuk SM, Senjem ML, White WM, Fields JA et al.
Influence of preeclampsia and late-life hypertension on MRI measures of cortical
atrophy. *Journal of Hypertension.* 2017;35:2479-85.
588 doi:10.1097/HJH.0000000000001492.
- 590 24. Brewer J, Owens MY, Wallace K, Reeves AA, Morris R, Khan M et al. Posterior
reversible encephalopathy syndrome in 46 of 47 patients with eclampsia. *Am J
Obstet Gynecol.* 2013;208(6):468 e1-6. doi:10.1016/j.ajog.2013.02.015.
- 592 25. Mielke MM, Milic NM, Weissgerber TL, White WM, Kantarci K, Mosley TH et al.
Impaired cognition and brain atrophy decades after hypertensive pregnancy
594 disorders. *Circulation: Cardiovascular Quality and Outcomes.* 2016;9:S70-S6.
doi:10.1161/CIRCOUTCOMES.115.002461.
- 596 26. Siepmann T, Boardman H, Bilderbeck A, Griffanti L, Kenworthy Y, Zwager C et al.
Long-term cerebral white and gray matter changes after preeclampsia. *Neurology.*
598 2017;88:1256-64. doi:10.1212/WNL.0000000000003765.
- 600 27. Mielke MM, Milic NM, Weissgerber TL, White WM, Kantarci K, Mosley TH et al.
Impaired Cognition and Brain Atrophy Decades After Hypertensive Pregnancy
602 Disorders. *Circ Cardiovasc Qual Outcomes.* 2016;9(1):S70-6.
doi:10.1161/CIRCOUTCOMES.115.002461.
- 604 28. Elharram M, Dayan N, Kaur A, Landry T, Pilote L. Long-Term Cognitive Impairment
After Preeclampsia: A Systematic Review and Meta-analysis. *Obstet Gynecol.*
2018;132(2):355-64. doi:10.1097/AOG.0000000000002686.
- 606 29. Cipolla MJ. Cerebrovascular function in pregnancy and eclampsia. *Hypertension.*
2007;50:14-24. doi:10.1161/HYPERTENSIONAHA.106.079442.

- 608 30. Johnson AC, Nagle KJ, Tremble SM, Cipolla MJ. The Contribution of Normal
Pregnancy to Eclampsia. *PLoS One*. 2015;10(7):e0133953.
610 doi:10.1371/journal.pone.0133953.
- 612 31. Cipolla MJ, Kraig RP. Seizures in Women with Preeclampsia: Mechanisms and
Management. *Fetal Matern Med Rev*. 2011;22(2):91-108.
doi:10.1017/S0965539511000040.
- 614 32. Coughlin WF, McMurdo SK, Reeves T. MR imaging of postpartum cortical blindness.
J Comput Assist Tomogr. 1989;13(4):572-6.
- 616 33. Trommer BL, Homer D, Mikhael MA. Cerebral vasospasm and eclampsia. *Stroke*.
1988;19(3):326-9.
- 618 34. Busija DW, Heistad DD. Factors Involved in the Physiological Regulation of the
Cerebral Circulation. *Reviews of Physiology Biochemistry and Pharmacology*.
620 1984;101:161-211.
- 622 35. Donaldson JO. Eclamptic hypertensive encephalopathy. *Seminars in Neurology*.
1988;8:230-3. doi:10.1055/s-2008-1041383.
- 624 36. Zeeman GG, Cipolla MJ, Cunningham FG. Cerebrovascular (Patho)Physiology in
Preeclampsia/Eclampsia. *Chesley's Hypertensive Disorders in Pregnancy*.
2009:227-47. doi:10.1016/B978-0-12-374213-1.00013-6.
- 626 37. Aagaard-Tillery KM, Belfort MA. Eclampsia: Morbidity, mortality, and management.
Clinical Obstetrics and Gynecology. 2005;48:12-23.
628 doi:10.1097/01.grf.0000153882.58132.ba.
- 630 38. van Veen TR, Panerai RB, Haeri S, Griffioen AC, Zeeman GG, Belfort MA. Cerebral
Autoregulation in Normal Pregnancy and Preeclampsia. *Obstetrics & Gynecology*.
2013;122:1064-9. doi:10.1097/AOG.0b013e3182a93fb5.
- 632 39. Brunner H, Cockcroft JR, Deanfield J, Donald A, Ferrannini E, Halcox J et al.
Endothelial function and dysfunction. Part II: Association with cardiovascular risk
634 factors and diseases. A statement by the Working Group on Endothelins and
Endothelial Factors of the European Society of Hypertension. *Journal of*
636 *hypertension*. 2005;23(2):233-46.
- 638 40. Roberts JM. Endothelial dysfunction in preeclampsia. *Seminars in reproductive*
endocrinology. 1998;16(1):5-15. doi:10.1055/s-2007-1016248.
- 640 41. Abbott NJ. Evidence for bulk flow of brain interstitial fluid: Significance for
physiology and pathology. *Neurochemistry International*. 2004;45:545-52.
doi:10.1016/j.neuint.2003.11.006.
- 642 42. Haseloff RF, Dithmer S, Winkler L, Wolburg H, Blasig IE. Transmembrane proteins of
the tight junctions at the blood-brain barrier: Structural and functional aspects.
644 *Seminars in Cell and Developmental Biology*. 2015;38:16-25.
doi:10.1016/j.semcdb.2014.11.004.
- 646 43. Keaney J, Campbell M. The dynamic blood-brain barrier. *FEBS Journal*.
2015;282:4067-79. doi:10.1111/febs.13412.
- 648 44. Daneman R, Prat A. The Blood - Brain Barrier. *Developmental Medicine & Child*
Neurology. 2015;3:311-4. doi:10.1111/j.1469-8749.1961.tb15323.x.
- 650 45. Decleves X, Jacob A, Yousif S, Shawahna R, Potin S, Scherrmann J-M. Interplay of
Drug Metabolizing CYP450 Enzymes and ABC Transporters in the Blood-Brain
652 Barrier. *Current Drug Metabolism*. 2011;12:732-41.
doi:10.2174/138920011798357024.

- 654 46. Liao MZ, Gao C, Shireman LM, Phillips B, Risler LJ, Neradugomma NK et al. P-
656 gp/ABCB1 exerts differential impacts on brain and fetal exposure to
norbuprenorphine. *Pharmacological Research*. 2017;119:61-71.
doi:10.1016/j.phrs.2017.01.018.
- 658 47. Aagaard-Tillery KM, Belfort MA. Eclampsia: morbidity, mortality, and management.
Clin Obstet Gynecol. 2005;48(1):12-23.
- 660 48. Warrington JP, Fan F, Murphy SR, Roman RJ, Drummond HA, Granger JP et al.
662 Placental ischemia in pregnant rats impairs cerebral blood flow autoregulation and
increases blood-brain barrier permeability. *Physiol Rep*. 2014;2(8).
doi:10.14814/phy2.12134.
- 664 49. Clayton AM, Shao Q, Paauw ND, Giambrone AB, Granger JP, Warrington JP.
666 Postpartum increases in cerebral edema and inflammation in response to placental
ischemia during pregnancy. *Brain Behav Immun*. 2018;70:376-89.
doi:10.1016/j.bbi.2018.03.028.
- 668 50. Black KD, Horowitz JA. Inflammatory Markers and Preeclampsia: A Systematic
Review. *Nursing Research*. 2018;67:242-51. doi:10.1097/NNR.000000000000285.
- 670 51. LaMarca BD, Ryan MJ, Gilbert JS, Murphy SR, Granger JP. Inflammatory cytokines in
672 the pathophysiology of hypertension during preeclampsia. *Current Hypertension
Reports*. 2007;9:480-5. doi:10.1007/s11906-007-0088-1.
- 674 52. Warrington JP, Drummond HA, Granger JP, Ryan MJ. Placental Ischemia-induced
Increases in Brain Water Content and Cerebrovascular Permeability: Role of TNF α .
676 *American journal of physiology Regulatory, integrative and comparative
physiology*. 2015:ajpregu.00372.2015. doi:10.1152/ajpregu.00372.2015.
- 678 53. Amburgey OA, Chapman AC, May V, Bernstein IM, Cipolla MJ. Plasma from
680 preeclamptic women increases blood-brain barrier permeability: role of vascular
endothelial growth factor signaling. *Hypertension*. 2010;56(5):1003-8.
doi:10.1161/HYPERTENSIONAHA.110.158931.
- 682 54. Li X, Han X, Bao J, Liu Y, Ye A, Thakur M et al. Nicotine increases eclampsia-like
684 seizure threshold and attenuates microglial activity in rat hippocampus through the
alpha7 nicotinic acetylcholine receptor. *Brain Res*. 2016;1642:487-96.
doi:10.1016/j.brainres.2016.04.043.
- 686 55. Huang Q, Liu L, Hu B, Di X, Brennecke SP, Liu H. Decreased seizure threshold in an
688 eclampsia-like model induced in pregnant rats with lipopolysaccharide and
pentylene tetrazol treatments. *PLoS One*. 2014;9(2):e89333.
doi:10.1371/journal.pone.0089333.
- 690 56. Eigenmann DE, Xue G, Kim KS, Moses AV, Hamburger M, Oufir M. Comparative
692 study of four immortalized human brain capillary endothelial cell lines,
hCMEC/D3, hBMEC, TY10, and BB19, and optimization of culture conditions, for
an in vitro blood-brain barrier model for drug permeability studies. *Fluids and
Barriers of the CNS*. 2013:1-17.
- 694 57. Rahman NA, Rasil ANaHM, Meyding-Lamade U, Craemer EM, Diah S, Tuah AA et
696 al. Immortalized endothelial cell lines for in vitro blood-brain barrier models: A
systematic review. *Brain Research*. 2016;1642:532-45.
doi:10.1016/j.brainres.2016.04.024.
- 698 58. Dauchy S, Dutheil F, Weaver RJ, Chassoux F, Dumas-Duport C, Couraud PO et al.
ABC transporters, cytochromes P450 and their main transcription factors:

- 700 expression at the human blood-brain barrier. *J Neurochem.* 2008;107(6):1518-28.
doi:JNC5720 [pii]
- 702 10.1111/j.1471-4159.2008.05720.x.
- 704 59. Dutheil F, Jacob A, Dauchy S, Beaune P, Scherrmann JM, Declèves X et al. ABC
transporters and cytochromes P450 in the human central nervous system: influence
706 on brain pharmacokinetics and contribution to neurodegenerative disorders. *Expert
Opin Drug Metab Toxicol.* 2010;6(10):1161-74.
doi:10.1517/17425255.2010.510832.
- 708 60. Ohtsuki S, Ikeda C, Uchida Y, Sakamoto Y, Miller F, Glacial F et al. Quantitative
targeted absolute proteomic analysis of transporters, receptors and junction proteins
710 for validation of human cerebral microvascular endothelial cell line hCMEC/D3 as a
human blood-brain barrier model. *Mol Pharm.* 2013;10(1):289-96.
712 doi:10.1021/mp3004308.
- 714 61. Bosworth AM, Faley SL, Bellan LM, Lippmann ES. Modeling Neurovascular
Disorders and Therapeutic Outcomes with Human-Induced Pluripotent Stem Cells.
Front Bioeng Biotechnol. 2017;5:87. doi:10.3389/fbioe.2017.00087.
- 716 62. Nielsen SSE, Siupka P, Georgian A, Preston JE, Tóth AE, Yusof SR et al. Improved
Method for the Establishment of an *In Vitro* Blood-Brain
718 Barrier Model Based on Porcine Brain Endothelial Cells. *Journal of Visualized
Experiments.* 2017. doi:10.3791/56277.
- 720 63. Smith M, Omid Y, Gumbleton M. Primary porcine brain microvascular endothelial
cells: Biochemical and functional characterisation as a model for drug transport and
722 targeting. *Journal of Drug Targeting.* 2007;15:253-68.
doi:10.1080/10611860701288539.
- 724 64. Zhang Y, Li CSW, Ye Y, Johnson K, Poe J, Johnson S et al. Porcine Brain Microvessel
Endothelial Cells as an *In Vitro* Model to Predict *In Vivo* Blood-Brain Barrier
726 Permeability. *DRUG METABOLISM AND DISPOSITION.* 2006;34:1-15.
doi:10.1124/dmd.105.006437.
- 728 65. Culot M, Lundquist S, Vanuxeem D, Nion S, Landry C, Delplace Y et al. An *in vitro*
blood-brain barrier model for high throughput (HTS) toxicological screening.
730 *Toxicol In Vitro.* 2008;22(3):799-811. doi:10.1016/j.tiv.2007.12.016.
- 732 66. Helms HC, Hersom M, Kuhlmann LB, Badolo L, Nielsen CU, Brodin B. An
electrically tight *in vitro* blood-brain barrier model displays net brain-to-blood
734 efflux of substrates for the ABC transporters, P-gp, Bcrp and Mrp-1. *AAPS J.*
2014;16(5):1046-55. doi:10.1208/s12248-014-9628-1.
- 736 67. Thomsen LB, Burkhart A, Moos T. A Triple Culture Model of the Blood-Brain Barrier
Using Porcine Brain Endothelial cells, Astrocytes and Pericytes. *PLOS ONE.*
2015;10:e0134765. doi:10.1371/journal.pone.0134765.
- 738 68. Helms HC, Waagepetersen HS, Nielsen CU, Brodin B. Paracellular tightness and
claudin-5 expression is increased in the BCEC/astrocyte blood-brain barrier model
740 by increasing media buffer capacity during growth. *AAPS J.* 2010;12(4):759-70.
doi:10.1208/s12248-010-9237-6.
- 742 69. Torres-Vergara P, Penny J. Pro-inflammatory and anti-inflammatory compounds exert
similar effects on P-glycoprotein in blood-brain barrier endothelial cells. *J Pharm
744 Pharmacol.* 2018;70(6):713-22. doi:10.1111/jphp.12893.

70. Salmeri M, Motta C, Anfuso CD, Amodeo A, Scalia M, Toscano MA et al. VEGF
746 receptor-1 involvement in pericyte loss induced by Escherichia coli in an in vitro
748 model of blood brain barrier. *Cell Microbiol.* 2013;15(8):1367-84.
doi:10.1111/cmi.12121.
71. Perriere N, Yousif S, Cazaubon S, Chaverot N, Bourasset F, Cisternino S et al. A
750 functional in vitro model of rat blood-brain barrier for molecular analysis of efflux
transporters. *Brain Res.* 2007;1150:1-13. doi:10.1016/j.brainres.2007.02.091.
72. Forster C, Silwedel C, Golenhofen N, Burek M, Kietz S, Mankertz J et al. Occludin as
752 direct target for glucocorticoid-induced improvement of blood-brain barrier
754 properties in a murine in vitro system. *J Physiol.* 2005;565(Pt 2):475-86.
doi:10.1113/jphysiol.2005.084038.
73. Alms D, Fedrowitz M, Romermann K, Noack A, Loscher W. Marked Differences in the
756 Effect of Antiepileptic and Cytostatic Drugs on the Functionality of P-Glycoprotein
758 in Human and Rat Brain Capillary Endothelial Cell Lines. *Pharm Res.*
2014;31(6):1588 - 604. doi:10.1007/s11095-013-1264-4.
74. Neuhaus W, Stessl M, Strizsik E, Bennani-Baiti B, Wirth M, Toegel S et al. Blood-
760 brain barrier cell line PBMEC/C1-2 possesses functionally active P-glycoprotein.
762 *Neurosci Lett.* 2010;469(2):224-8. doi:S0304-3940(09)01570-5 [pii]
10.1016/j.neulet.2009.11.079.
75. Neuhaus W, Plattner VE, Wirth M, Germann B, Lachmann B, Gabor F et al. Validation
764 of in vitro cell culture models of the blood-brain barrier: tightness characterization
766 of two promising cell lines. *J Pharm Sci.* 2008;97(12):5158-75.
doi:10.1002/jps.21371.
76. Belfort MA, Saade GR, Yared M, Grunewald C, Herd JA, Varner MA et al. Change in
768 estimated cerebral perfusion pressure after treatment with nimodipine or magnesium
770 sulfate in patients with preeclampsia. *American Journal of Obstetrics and
Gynecology.* 1999;181:402-7. doi:10.1016/S0002-9378(99)70569-7.
77. Belfort MA, Tooke-Miller C, Allen JC, Dizon-Townson D, Varner MA. Labetalol
772 decreases cerebral perfusion pressure without negatively affecting cerebral blood
774 flow in hypertensive gravidas. *Hypertension in Pregnancy.* 2002;21:185-97.
doi:10.1081/PRG-120015845.
78. Belfort MA, Varner MW, Dizon-Townson DS, Grunewald C, Nisell H. Cerebral
776 perfusion pressure, and not cerebral blood flow, may be the critical determinant of
778 intracranial injury in preeclampsia: A new hypothesis. *American Journal of
Obstetrics and Gynecology.* 2002;187:626-34. doi:10.1067/mob.2002.125241.
79. Van Veen TR, Panerai RB, Haeri S, Singh J, Adusumalli JA, Zeeman GG et al.
780 Cerebral autoregulation in different hypertensive disorders of pregnancy. *American
782 Journal of Obstetrics and Gynecology.* 2015;212:513.e1-e7.
doi:10.1016/j.ajog.2014.11.003.
80. Valdueza JM, Balzer JO, Villringer A, Vogl TJ, Kutter R, Einhaupl KM. Changes in
784 blood flow velocity and diameter of the middle cerebral artery during
786 hyperventilation: assessment with MR and transcranial Doppler sonography. *AJNR
Am J Neuroradiol.* 1997;18(10):1929-34.
81. Williams DS, Detre JA, Leigh JS, Koretsky AP. Magnetic resonance imaging of
788 perfusion using spin inversion of arterial water. *Proc Natl Acad Sci U S A.*
790 1992;89(1):212-6.

- 792 82. Le Bihan D. Intravoxel incoherent motion imaging using steady-state free precession.
Magn Reson Med. 1988;7(3):346-51.
- 794 83. Nelander M, Hannsberger D, Sundstrom-Poromaa I, Bergman L, Weis J, Akerud H et
al. Assessment of cerebral perfusion and edema in preeclampsia with intravoxel
796 incoherent motion MRI. Acta Obstet Gynecol Scand. 2018;97(10):1212-8.
doi:10.1111/aogs.13383.
- 798 84. Jansen JF, Backes WH, Nicolay K, Kooi ME. 1H MR spectroscopy of the brain:
absolute quantification of metabolites. Radiology. 2006;240(2):318-32.
doi:10.1148/radiol.2402050314.
- 800 85. Oberhaensli RD, Galloway GJ, Hilton-Jones D, Bore PJ, Styles P, Rajagopalan B et al.
802 The study of human organs by phosphorus-31 topical magnetic resonance
spectroscopy. Br J Radiol. 1987;60(712):367-73. doi:10.1259/0007-1285-60-712-
367.
- 804 86. Nelander M, Weis J, Bergman L, Larsson A, Wikstrom AK, Wikstrom J. Cerebral
806 Magnesium Levels in Preeclampsia; A Phosphorus Magnetic Resonance
Spectroscopy Study. American journal of hypertension. 2017;30(7):667-72.
doi:10.1093/ajh/hpx022.
- 808 87. Nelander M, Wikstrom AK, Weis J, Bergman L, Larsson A, Sundstrom-Poromaa I et
810 al. Cerebral Osmolytes and Plasma Osmolality in Pregnancy and Preeclampsia: A
Proton Magnetic Resonance Spectroscopy Study. Am J Hypertens. 2018;31(7):847-
53. doi:10.1093/ajh/hpy019.
- 812 88. Rutherford JM, Moody A, Crawshaw S, Rubin PC. Magnetic resonance spectroscopy in
pre-eclampsia: evidence of cerebral ischaemia. BJOG. 2003;110(4):416-23.
- 814 89. Sengar AR, Gupta RK, Dhanuka AK, Roy R, Das K. MR imaging, MR angiography,
816 and MR spectroscopy of the brain in eclampsia. AJNR American journal of
neuroradiology. 1997;18(8):1485-90.
- 818 90. Marchi N, Cavaglia M, Fazio V, Bhudia S, Hallene K, Janigro D. Peripheral markers of
blood-brain barrier damage. Clin Chim Acta. 2004;342(1-2):1-12.
doi:10.1016/j.cccn.2003.12.008.
- 820 91. Kanner AA, Marchi N, Fazio V, Mayberg MR, Koltz MT, Siomin V et al. Serum
822 S100 β : A noninvasive marker of blood-brain barrier function and brain lesions.
Cancer. 2003;97:2806-13. doi:10.1002/cncr.11409.
- 824 92. Schmidt A, Tort A, Amaral O, Schmidt A, Walz R, Vettorazzi-Stuckzynski J et al.
Serum S100B in Pregnancy-Related Hypertensive Dis- orders: A Case-Control
826 Study. Clinical Chemistry. 2004;50:435-8. doi:10.1373/clinchem.2003.027391.
- 828 93. Vettorazzi J, Torres FV, de Avila TT, Martins-Costa SH, Souza DO, Portela LV et al.
Serum S100B in pregnancy complicated by preeclampsia: A case-control study.
Pregnancy Hypertens. 2012;2(2):101-5. doi:10.1016/j.preghy.2011.11.004.
- 830 94. Bergman L, Akhter T, Wikstrom AK, Wikstrom J, Naessen T, Akerud H. Plasma levels
of S100B in preeclampsia and association with possible central nervous system
832 effects. Am J Hypertens. 2014;27(8):1105-11. doi:10.1093/ajh/hpu020.
- 834 95. Artunc-Ulkumen B, Guvenc Y, Goker A, Gozukara C. Maternal Serum S100-B, PAPP-
A and IL-6 levels in severe preeclampsia. Archives of Gynecology and Obstetrics.
2015;292:97-102. doi:10.1007/s00404-014-3610-0.
- 836 96. Chou SHY, Robertson CS. Monitoring Biomarkers of Cellular Injury and Death in
Acute Brain Injury. Neurocritical Care. 2014;21:187-214. doi:10.1007/s12028-014-
0039-z.

- 838 97. Bergman L, Akerud H. Plasma Levels of the Cerebral Biomarker, Neuron-Specific
Enolase, are Elevated During Pregnancy in Women Developing Preeclampsia.
840 *Reprod Sci.* 2016;23(3):395-400. doi:10.1177/1933719115604732.
- 842 98. Bergman L, Akerud H, Wikström AK, Larsson M, Naessen T, Akhter T. Cerebral
biomarkers in women with preeclampsia are still elevated 1 year postpartum.
American Journal of Hypertension. 2016;29:1374-9. doi:10.1093/ajh/hpw097.
- 844 99. Bogoslovsky T, Gill J, Jeromin A, Davis C, Diaz-Arrastia R. Fluid biomarkers of
traumatic brain injury and intended context of use. *Diagnostics.* 2016;6:1-22.
846 doi:10.3390/diagnostics6040037.
- 848 100. Randall J, Mörtberg E, Provuncher GK, Fournier DR, Duffy DC, Rubertsson S et al.
Tau proteins in serum predict neurological outcome after hypoxic brain injury from
cardiac arrest: Results of a pilot study. *Resuscitation.* 2013;84:351-6.
850 doi:10.1016/j.resuscitation.2012.07.027.
- 852 101. Evers KS, Atkinson A, Barro C, Fisch U, Pfister M, Huhn EA et al. Neurofilament as
Neuronal Injury Blood Marker in Preeclampsia. *Hypertension.* 2018;71(6):1178-84.
doi:10.1161/HYPERTENSIONAHA.117.10314.
- 854 102. Bergman L, Zetterberg H, Kaihola H, Hagberg H, Blennow K, Akerud H. Blood-based
cerebral biomarkers in preeclampsia: Plasma concentrations of NfL, tau, S100B and
856 NSE during pregnancy in women who later develop preeclampsia - A nested case
control study. *PLoS One.* 2018;13(5):e0196025. doi:10.1371/journal.pone.0196025.

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